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Stem Cells in Head & Neck Medicine

Update on the Role of Emerging Stem Cell Technology in Head and Neck Medicine

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Abstract

Head and neck surgery is a broad discipline that involves the management of complex conditions such as: burns, skin cancer, head and neck cancer, congenital abnormalities and facial rejuvenation. For cancer patient's; surgery, radiotherapy and chemotherapy are often the main modes of treatment. Many patients require follow up reconstructive surgery, and the use of stem cells offers novel treatments that could aid recovery. Laryngeal, tracheal and neuronal tissues are frequently damaged by surgery in the head and neck and these tissues have little intrinsic regenerative ability. Pluripotent embryonic stem cells retain the ability to differentiate into a wide variety of cells meaning that large tissue defects can be reduced by stimulating new cell growth. Research has demonstrated potential benefits of using stem cells in facial rejuvenation procedures and the management of burns sequelae. The advancements made in the use of adult progenitor stem cells as a possible source for pluripotent stem cells (Induced pluripotent stem cells) mean that ethical considerations around the use of embryological tissue can be minimised, allowing for more research to take place. Currently the evidence base for the use of stem cells in head and neck surgery is limited, but it has now been proven that stem cells can act as a source for lost or damaged tissue in the head and neck. With continuous advancements being made in the fields of tissue engineering; it is likely that stem cells will play a major role in head and neck surgery in the future.

Keywords

Regenerative Medicine, Mesenchymal Stem Cells, Biomaterials, Scaffolds, Head and Neck Surgery

Introduction

Stem cells are undifferentiated cells that have the unique properties of differentiating into a variety of cell types and self-renewal. Embryonic stem cells (ESCs) are found on the inner wall of the blastocyst in the early embryo¹. ESCs are the primary example of pluripotent stem cells as they are able to self-renew and differentiate into the three-germ layers: ectoderm, endoderm and mesoderm, but are incapable of forming placental tissue (Figure 1). Adult stem cells are multipotent stem/progenitor cells that are able to differentiate into specific cell lineages. Mesenchymal stem cells (MSCs) are an example of adult stem cells². MSCs possess the ability to be altered to resemble pluripotent embryonic stem cells in both their morphology and growth properties by the introduction of transcription factors, by means of transfection of defined genes such as POU5F1 (POU Class 5 Homeobox 1), SOX2 (SRY (sex determining region Y) -box2), Klf4 (Kruppel-like factor-2) and cMyC (Master Regulator of cell cycle and Proliferative metabolism)³. These transformed adult progenitor cells are referred to as induced pluripotent stem cells (iPSCs). By using iPSCs as a source of pluripotent stem cells it is possible to minimise the ethical concerns over the use of embryonic tissue, allowing more research into the field of stem cell therapy to be undertaken. The iPSCs have been shown to express ESC markers such as: Stage Specific Embryonic Antigen-3 (SSEA-3), Stage Transformer-2 protein homolog (TRA-2-29/6E) and NANOG, whilst also expressing genes which are observed in human ESC such as: POU class 5, transcription factor 1 (POU5F1), SRY-Box Transcription factor-2 (SOX2), NANOG, Growth differentiation factor-3 (GDF3), Fibroblastic Growth Factor-4 (FGF4), Development Pluripotency Associated-2 (DPPA2), Development Pluripotency Associated-4 (DPPA4), human Telomerase reverse transcriptase (hTERT) and REX1 transcription factor (REX1), although the genes expressed show distinct expression level at different passages during expansion⁴. One major concern with the use of both iPSCs and ESCs is the risk of teratoma formation, which should be considered when using both ESCs and iPSCs⁵.

Stem Cells in Head & Neck Medicine

Head and neck surgery often involves removal of significant amounts of tissue, this is particularly true of the surgical management of head and neck cancers. These operations often require further reconstructive surgeries that use numerous tissues with poor regenerative capabilities to restore both form and function to the patient. Cartilage is an avascular tissue with no intrinsic regenerative capabilities and therefore large postoperative defects in a patient's cartilage significantly impede their recovery. MSCs are progenitor cells of mesodermal origin that have the ability to differentiate into chondrocytes, osteocytes and adipocytes². MSCs have been isolated from an ever-expanding array of mesodermal tissues and have been shown to express a range of cell surface markers. The most common set of MSC markers have been set out by the International Society for stem cell research (ISSCR) as the minimal criteria (Table 1)⁶. However, classification is not limited to the minimal criteria set out by the ISSCR (Table 2)⁷. MSCs ability to differentiate into chondrocytes whilst retaining their own self renewing properties means they have the potential to help regeneration of cartilage defects following reconstructive surgery. Animal models have already shown that intra articular injections of MSCs are able to help regenerate articular cartilage defects⁸. However, MSCs regenerative capabilities are not limited to chondrocytes, MSC's have been shown to be capable of differentiation into a wide variety of cell types within the ex-vivo setting (Figure 2)⁹. It has also been demonstrated that MSCs can influence fibroblasts and wound healing through paracrine effects of cytokines and chemokines¹⁰. This ability to educate and influence the wound environment makes MSCs an attractive adjunct therapy in treating burns wounds. Mesenchymal stem cells can be isolated easily from adipose tissue, bone marrow and umbilical cord meaning they are not subject to ethical controversy surrounding the use of embryonic tissue¹¹. MSCs have also been shown to have minimal alloreactivity which reduces the risk of allogeneic rejection¹². It has also been demonstrated in mouse animal models that mice treated with MSCs derived from bone marrow saw significantly better wound healing coverage following escharotomy than those that were untreated¹³. Stromal vascular fraction (SVF), which contains adipose and mesenchymal stem cells, has also been found to improve the healing of burn

Stem Cells in Head & Neck Medicine

wounds by increasing proliferation, vascularisation and fibroblast activity whilst simultaneously reducing inflammation in the wound bed ¹⁴.

Unlike iPSCs, MSC's do not pose a risk of teratoma formation, which remains a major risk factor in the use of patient derived iPSCs. The capacity of stem cell use in both reconstructive surgery and tissue regeneration continues to be a multi-disciplinary area. Stem cells alone are not a fix all, golden bullet. Currently, Stem cells are utilised in a multi-component system called tissue engineering with a goal to produce a favourable outcome for the patient. Tissue engineering is best described as controlled differentiation by means of stimuli. This differentiation may be delivered by means of a scaffold system, which may or may not contain an array of chemical stimulants capable of inducing differentiation towards the tissue of interest. Furthermore, scaffolds should be capable of delivering mechanical stimulus to the naïve or differentiating cells to further guide the cells in a process known as mechanotransduction induced differentiation (Figure 3).

The use of stem cells in tissue engineering as shown in Figure 2 shows the intricate interactions between many factors such as; mechanical stimulation, scaffold/substrate stiffness, choice of stem cells, and the level of oxygen saturation. Taking all these factors into account would lead to a more suitable implant being engineered for use in patients.

Tissue Engineering

Tissue engineering was first coined in the 1988 National Science Foundation workshop. The concept of tissue engineering was championed by Caplan in the late 1980's ^{15, 16} with Caplan's work mainly focusing on the use of mesenchymal stem cells and the ability of MSCs to transdifferentiate between Osteogenic, chondrogenic and adipogenic phenotypes under the correct biochemical and growth factor cues ^{15, 17, 18}. Following this early research many other disciplines such as polymer sciences and drug release research started to collaborate in means of adding structural support and delivery

Stem Cells in Head & Neck Medicine

methods for stem cells to aid further in tissue engineering with a main goal of delivering a functional replacement tissue.

Biomaterials/ Bio-scaffolds

The concept of biomaterials was first proposed at the 1976 Consensus conference in the European society for biomaterials (ESB) and defined as “a nonviable material used in a medical device, intended to interact with a biological systems”¹⁹. Currently, the ESB has adapted the definition to: “material intended to interface with biological systems to evaluate, treat, augment or replace any tissue organ or function of the body”¹⁹. This adaptation highlights the progression in complexity of current biomaterials.

Currently, there are a wide range of biomaterials available for use in tissue engineering. These biomaterials can range from: ceramics such as hydroxyapatite (HA) and tri-calcium phosphate (TCP); which are used in bone regeneration^{20,21}, Poly(L-lactide-co-glycolide) PLGA²², Poly-L-lactic acid (PLLA)²³, Poly (lactide) PLA;²⁴ used in chondrogenesis or even co-polymers such as poly (3-hydroxybutyrate-co-3-hydroxyhexanoate) PHBHHx used in tendon repair²⁵. There are at least 3 major attributes to a biomaterial which will be used as a bio-scaffold. These are: Biocompatibility, Biodegradability and Mechanical properties^{19, 26, 27}. In regards biocompatibility, the implant should not induce an immune response as a complete implant or when biodegrading. Preferably there should be an inverse relationship between the rate of degradation and tissue regeneration. Ideally, prior to implantation the scaffold/implant should be able to withstand the natural forces exposed to the target tissue being replaced, which would then prevent implant failure as seen in Webb et al 2013 where all components of the implant were mechanically tested to produce the optimum scaffold for implantation²⁵.

Mechanical stimulation

Stem Cells in Head & Neck Medicine

Mesenchymal stem cells and progenitors have been shown to have distinctively different mechanical properties during their expansion in cell culture when analysed using atomic force microscopy (AFM) ^{28, 29}. Biomaterial based scaffolds have been used in-vivo to induce mechanical stimulation which has been shown to drive stem cell differentiation towards a target tissue. Osteogenesis has been shown to be induced by cyclic-compression loading ³⁰, whilst shear-stress loading by means of flow-perfusion bioreactors has been shown to induce chondrogenic differentiation ³¹. Implanted acellular scaffolds have also demonstrated cellular invasion into the scaffold core which comprised rat-tail collagen and cellular alignment along the force bearing fibres in tendon implants ²⁵. This further demonstrates the mechano-responsive capability of both mesenchymal, progenitor and tissue specific cell types. Another important possible result of mechanical loading is that during mechanical loading the extra-cellular fluid within the regenerating tissue is forced out taking waste metabolites away from the cells. When the force is relaxed fresh fluid enters the extra-cellular space replenishing cellular nutrients and oxygen (mechanical loading induced perfusion) ³².

Growth factors

Growth factors are peptide based chemical signals that are essential in embryonic and adult tissue development. The major groups of growth factors used in tissue engineering are the transforming growth factor (TGF) superfamily of growth factors, which include Bone morphogenetic proteins (BMPs), and the Fibroblastic Growth factor (FGF) family. The over expression of members of the TGF superfamily have been linked to many malignancies including breast and ovarian cancers ³³. Over expression of FGF has also been shown to be linked with malignancies ³⁴. Therefore, during growth factor induced differentiation there must be careful consideration when selecting the concentration of growth factors used and exposure time. Most growth factor induced differentiation results in the up-regulation of transcription factors responsible for differentiation towards a cell lineage. TGF β ₁ results in up-regulation of SOX9 transcription factor in chondrogenesis via the SMAD1/5/8 signal

Stem Cells in Head & Neck Medicine

cascade³⁵, whilst BMP9 up-regulates RUNX2 transcription factor for bone differentiation via SMAD1/5/8 and a heterodimeric complex SMAD4 within the cell nucleus during differentiation³⁶. Conversely, BMP9 has more recently been utilised to differentiate chondroprogenitors towards a chondrocyte phenotype using 100ng/ml for 14 days. This resulted in a thin layer of cartilage when compared to traditional TGFβ₁ differentiation using pellet culture, which may indicate maturation or an osteoarthritic disease model³⁷. Although growth factors initiate transcription factor expression, other co-factors such as: culture media components, mechanical stimulation, cell density and oxygen tension should be considered during the differentiation and maturation of the engineered tissue.

Oxygen Tension

Oxygen tension has been shown to significantly influence differentiation³⁸. Most cell culture is conducted at 21% O₂ environment, but alveolar epithelial cells are exposed to O₂ levels between 13 - 14%, adipose tissue is exposed to O₂ levels between 3 - 8%, bone is exposed to O₂ levels between 2 - 5% and cartilage is exposed to O₂ levels between 1.4 - 2%³⁹. Dale et al showed that embryonic stem cells could be differentiated towards a tendon lineage at an oxygen tension of 2% but not 21% using BMP12 and 13⁴⁰. Gomez-Leduc et al showed oxygen tension <5% reduced the presence of mature chondrocyte markers when compared to 21% O₂ differentiation using TGFβ₁⁴¹. Modulation and control of oxygen tension should not be overlooked when tissue engineering a construct for use in research or patient implantation.

Head and Neck Surgery

Head and neck surgery remains a broad discipline involving a wide range of surgical techniques. Surgery of the head and neck involves the management of congenital abnormalities, trauma, burns and malignancy. Head and neck malignancies continue to be very challenging to treat. Surgical

Stem Cells in Head & Neck Medicine

removal often remains the gold standard treatment, followed by adjunct chemotherapy, radiotherapy and further reconstructive surgery. These surgeries often result in both large defects in structures such as trachea and vocal tissue and iatrogenic damage to neuronal tissue. Stem cells unique regenerative capabilities means they could potentially play a major role in treating these patients and restoring lost function when utilised in conjunction with tissue engineering.

Cranial Nerves

Nerves in the head and neck are often susceptible to damage during surgery. Damage to these nerves can result in areas of facial paralysis, sensory deficit and difficulties in phonation and swallowing, all of which can severely affect a patient's quality of life.

Recurrent Laryngeal Nerve

The recurrent laryngeal nerve is a branch of the vagus nerve that takes an indirect course through the neck supplying the intrinsic muscles of the larynx and sensation to the inferior portion of the larynx. Damage to the recurrent laryngeal nerve may result in vocal cord paralysis or hoarseness of the voice. A 20-year longitudinal study found that surgical procedures were the most common cause of injury to this nerve with thyroidectomy causing the majority (80%) of iatrogenic bilateral vocal fold immobility⁴². A recent study demonstrated the potential benefit of stem cells on returning function after transection of the recurrent laryngeal nerve. A group of 33 mice had their recurrent laryngeal nerve transected before being treated with a stem cell scaffold. Following the treatment 80% of the mice treated with the hybrid-transplantation of skeletal muscle-derived stem cells and bioabsorbable scaffold showed spontaneous movement of their vocal folds during breathing ⁴³. It has been demonstrated that laminin protein that is found in the extracellular matrix can play an important role in inducing embryonic and neuronal stem cells to differentiate and aid in the regeneration of nervous tissue⁴⁴. A study by Li Yu et al. found that by using a PLGA biomaterial scaffold embedded with laminin

Commented [HS1]: This section has been altered in response to the reviewers comment on neuronal tissue. We focused specifically on cranial nerves as a section on cervical nerves was also considered although the body of research to support this section was felt to be lacking. As a result the title has been changed as per the reviewers comments to reflect the focus on cranial nerves

Commented [HS2]: Sub-Headings have been included for the facial nerve and RLN sections and they have been split into two separate subsections

Stem Cells in Head & Neck Medicine

chitosan they were able to promote recurrent laryngeal nerve growth in a rat model ⁴⁵. By utilising the interplay between the biomaterials and the stem cells researchers are able to create microenvironments that allow stem cells to guide and sustain neuronal tissue growth. More recently Li Yu has also shown that the use of adipose-derived mesenchymal stem cells or differentiated Schwann-like adipose-derived mesenchymal stem cells result in improved healing and regeneration of the laryngeal nerve when compared to the use of extracellular matrix alone. When the mesenchymal stem cells were injected at the injury site a higher density of myelinated nerve fibre, thicker myelin sheath, improved vocal fold movement and better recovery of nerve conduction capacity were observed ⁴⁶. These results highlight the possible adjunct therapeutic advantage that stem cells could introduce when considering autologous tissue grafting.]

Facial Nerve

The facial nerve is one of the most common cranial nerve injuries following trauma to the head and neck, with facial paralysis occurring in 50% of cases of transverse fractures of the temporal bone ⁴⁷. Traumatic facial nerve injuries may result from transection, bony impingement on the nerve or intraneural hematoma ⁴⁷. These injuries can be partial such as crush injuries or they can involve complete transection of the nerve which poses an additional challenge as cut ends of the nerve often retract leaving a gap. Currently the options for surgical repair of these segmental nerve defects is limited. Nerve autografts are considered to be the gold standard treatment despite the fact that nerve autografts leave a donor site deficit and are therefore in limited supply ⁴⁸. Interestingly it has been demonstrated in a rabbit model of facial nerve injury that nerve conduits implanted with neural stem cells are not inferior to nerve autografts in terms of regeneration and repair. 10mm gaps in rabbit facial nerves were treated with chitosan conduit, collagen protein sponge, nerve growth factor (NGF), and neuronal stem cells (NSC). When compared with gaps treated by autograft or chitosan conduit and collagen sponge it was found that the nerves treated with NSC implanted conduits were thicker than the nerves treated with the autografts or conduit alone and electroneurophysiological detection

Commented [HS3]: This section has been expanded to include a greater volume of recent research that helps to demonstrate the potential advancements being made in the use of stem cells with biomaterial scaffolds.

Stem Cells in Head & Neck Medicine

showed no significant difference between groups ⁴⁹. This study demonstrates that there when comparing nerve autografts with NSC derived stem cells in a bio-scaffold there is no significant difference in nerve regeneration and that using conduits embedded with the appropriate stem cells may be equal to nerve autografts.

Although the studies above show promising results they are all animal models which limits their translation to human trials. However more recently small scale studies involving human subjects have begun to appear in the literature. A study of 8 patients who had all suffered traumatic transection of their facial nerve found significant clinical improvement in angulation of the mouth and closure of the eye as well as improvements in electroneuronography following injection of MSCs at the site of the nerve transection. Although these results are very promising the scale of the study is very small and patients were only followed up for 6-month time period, so it is unclear whether these benefits were maintained ⁵⁰.

Tracheal Tissue

Primary cancer of the trachea is quite rare, but it is associated with high mortality ^{51, 52}. The first paediatric, stem cell-based tissue engineered tracheal transplant undertaken in 2012 showed promising results ⁵³. However, more recently the use of stem cells in tracheal transplantation has been subject to a great deal of controversy as Paolo Macchiarini, a pioneer of the use of stem cell seeded tracheal transplants, has been accused of research misconduct and unethical experimental operations after a number of his patients died after receiving his transplants. This incident demonstrates the risks that experimental stem cell procedures could have for patients and highlights the need for closer regulation and monitoring in future trials. Current research into 3D printed nanofiber artificial trachea has demonstrated that by combining the artificial fibres with iPSC-derived MSCs and chondrocytes it is possible to create a functional epithelized, cartilaginous airway ⁵⁴. The success of this stem cell

Commented [HS4]: The section covering the facial nerve has been made into its own individual subheading and it has been greatly expanded with current research into animal models to demonstrate the potential benefit that could be achieved. The section towards the end describing early small scale work in human trials has been kept but moved to the end of the passage

Stem Cells in Head & Neck Medicine

infused artificial scaffold shows the importance of identifying populations of stem cells that have the capability to differentiate into cartilaginous tissue. Recent studies have highlighted a population of stem/progenitor cells that reside within the tracheal C-ring²⁸. The cell population expressed common MSC markers and were shown to be capable of tri-lineage differentiation. However, the isolated stem/progenitor cells were shown to undergo calcification when differentiated towards a chondrocyte lineage using the growth factors: Transforming Growth Factor Beta-1 (TGF β ₁) at a concentration of 10ng/mL or Bone Morphogenic Protein-9/Growth Differentiation Factor-5 (BMP9/GDF5) at concentration of 100ng/ml. This may suggest that further optimisation of differentiation components is required²⁸. Despite this researchers have gone on to show that isolated autologous bone marrow derived stem cells are able to differentiate into chondrocytes when exposed to TGF β ₁ in an in vivo transplantation of a genipin linked tracheal scaffold⁵⁵. Currently the concerns surrounding stem cell usage with tracheal tissue limit its application in clinical practice but it is possible that future studies may elucidate the potential of these cells for reconstructive surgery of the trachea.

Commented [HSS]: More papers have been included in the tracheal section to both highlight the stem cell work being done surrounding artificial tracheal structures and their similarities to human tissue when embedded with stem cells and also how stem cells have been induced to differentiate into chondrocytes under specific cell environments

Vocal Tissue

Surgical management of laryngeal cancer involves a partial or total laryngectomy. Partial laryngectomy will have an impact on voice quality and speech intelligibility regardless of surgical technique due to damage to the vocal folds⁵⁶. Human vocal fold epithelium has very little proliferative capacity and as a result it has been difficult to produce models that allow for more research into novel therapies. The use of stem cells could allow for regeneration of these damaged tissues as it has been demonstrated that a 3D model of vocal fold mucosa can be reliably created by using ESC-derived epithelial cells and human primary vocal fold fibroblasts⁵⁷. It has even been shown that human iPSCs can be modulated by FGF to produce a vocal fold mucosa that is able to mimic inflammatory response to smoke exposure seen in human vocal fold tissue⁵⁸. Reconstructive phonosurgery will also often lead to scarring of the vocal folds further worsening dysphonia in some patients. A recent systematic review of the use of

Commented [HS6]: This section has also been expanded to include more studies and a more in depth look at examples of recent and current research that demonstrate the potential of stem cell therapy better.

Stem Cells in Head & Neck Medicine

MSCs as a treatment for vocal fold scarring in pre-clinical studies found promising results, although it is difficult to assess whether these positive findings will be seen in human clinical trials⁵⁹. It is unclear exactly how MSCs interact with the vocal fold mucosa at a tissue level to minimise scarring but it has been demonstrated in a rat model that MSCs can suppress $TGF\beta_1$ signalling in vocal fold fibroblasts aiding their anti-fibrotic effect. Interestingly it was also noted that bidirectional paracrine signalling meant that the native vocal fold fibroblasts were also able to regulate the ECM in the MSCs⁶⁰. Similar positive effects on organisation of the ECM collagen matrix have been observed on animal models involving vocal fold resection. The lamina propria of rabbits vocal folds was resected and human MSCs were injected at the site of resection. Histological analysis demonstrated a reduced inflammatory response with increased levels of CD163+ anti-inflammatory macrophages and greater organisation in the collagen matrix⁶¹. Although much of the work investigating stem cells effects on vocal tissue healing has been completed in animal models early work in human clinical trials is also promising, showing improvements in phonation following treatment, although more large-scale research is needed to fully assess these claims⁶².

Soft Tissue Regeneration

As well as aiding in the regeneration and healing of neuronal and cartilaginous tissue, stem cells can play an important role in soft tissue regeneration in the head and neck.

Head and Neck Burns

The use of stem cells in burn patients is multifaceted as they can help to regenerate epidermal tissue, educate the wound environment and model the healing process to reduce fibrosis and scar formation. Currently autologous skin grafts are widely used but problems can arise when there is not a sufficient or appropriate donor site area⁶³. Burn injury sequelae such as hypertrophic scarring, contracture and

Commented [HS7]: This subsection has been changed to "Soft Tissue Regeneration" as suggested by the reviewer. It has now been split into 3 sub sections.

Commented [HS8]: This subsection has been reworked with more added to put a greater focus onto burns that affect the head and neck specifically.

Stem Cells in Head & Neck Medicine

pigmentation changes in the skin often require rehabilitation and physiotherapy ⁶⁴. This can produce unique challenges in the head and neck as scar contractures in the neck can have a negative impact on breathing, phonation and mastication and will often require reconstructive surgery. Adipose derived stem cells have been shown to improve chronic neck scar contracture and wound healing following radiation burns ^{65, 66}. Post burn scars in the face and neck can create a significant psychological burden for patients recovery. Nanofat injections containing stromal vascular fraction rich in adipose-derived stem cells have been shown to produce significantly improved scar appearance when injected at the site of none hypertrophic postburn scars in patients ⁶⁷. This research highlights that stem cells may play an important role in helping to improve postburn scarring in the face.

Facial Scarring

Hypertrophic scars are notoriously difficult to manage but recent work has suggested that MSCs may play a role in helping to prevent their formation. It has been shown in a mice model that when MSCs are combined with fibroblasts and introduced to the wound environment they can alter the fibroblast activity via paracrine effects preventing the formation of hypertrophic scar tissue ⁶⁸. The discovery of MSCs in adipose tissue has meant that autologous fat grafting is now being utilised more frequently in the treatment of facial scarring. A recent review demonstrated that clinical improvement in scars has been demonstrated in almost every area of the face, especially if scarring is derived from burns, trauma, degenerative diseases, and radiotherapy although the authors did raise concerns over the fact that there is considerable variability in outcome between different patients receiving the same treatment ⁶⁹. Despite this, autologous fat grafting has been shown to be as effective at treating acne scarring as established techniques such as fractional carbon dioxide laser (FxCr). It was shown that just one treatment using autologous adipose tissue-derived adult stem cells provided significant global improvement when compared to 3 treatments involving FxCr over a 3-month period ⁷⁰.

Commented [HS9]: This section now looks specifically at how soft tissue regeneration can be applied to scars in the face and what benefits stem cell therapy can provide.

Stem Cells in Head & Neck Medicine

Epidermal Stem Cells

Epidermal stem cells are found in both the epidermis and the hair follicle of the skin. Cultured epithelial autografts containing these epidermal stem cells have become an established treatment in burn injuries⁶³. Advancements in tissue engineering have meant that the epidermal stem cells have been combined with fibrin matrices in attempts to improve durability and to try and minimise rejection of the graft. As well as concerns over durability, cost and rejection of transplanted tissue, there are also concerns over mutation accumulation over several cell passages of cell division⁷¹. However, recently it has been shown that a limited number of transgenic epidermal stem cells were able to regenerate the entire epidermis, once again demonstrating the potential that these cells have in treating major burn injuries⁷². It is worth noting that the stem cells role in aiding the regeneration of the epidermis is not limited to the treatment of burn injuries. It is possible that stem cells epidermal regenerative capabilities could be applied to surgical defects in the head and neck to aid in regeneration of dermal tissue. Recent studies in animal models have demonstrated that tonsillar MSCs incorporated into wound beds significantly promoted the repair of surgical defects in mice with a considerable impact on the augmentation of epidermal and dermal regeneration⁷³. It has already been demonstrated in patients who have undergone surgery to their head and neck that higher levels of pro-inflammatory cytokines such as interleukin-1 β , interleukin-8, and Matrix metalloproteinase -9 are associated with both greater incidence of postoperative complications and length of stay in hospital⁷⁴. The paracrine factors released by MSCs are intricately linked with the modulation of the inflammatory environment and they play an important role in regulating levels of these pro inflammatory cytokines⁷³. By utilising stem cells in head and neck surgery it is possible that inflammatory response could be reduced, helping to improve postoperative outcomes and complications following surgery in the head and neck.

Commented [HS10]: Here the epidermal regenerative capabilities of stem cells have been linked directly to surgery in the head and neck as per the suggestion by the reviewers.

Discussion

Following the discovery of induced pluripotent stem cells and tissue specific reservoirs of adult stem cells i.e. Tracheal C-ring, there has been a wealth of research that has investigated stem cells as a novel therapy to treat all kinds of disease. Much of this work has looked at stem cells regenerative abilities with countless animal models demonstrating positive results. As a result, stem cells remain a promising therapeutic option for tissue regeneration and wound healing in the field of head and neck surgery. However, despite this wealth of information demonstrating the potential benefits of stem cells in all areas of head and neck surgery, there is a distinct lack of evidence in human trials. Very recently there has been an increase in the number of small-scale case reports showing that stem cell therapy can be a viable therapy in humans. Advancements in bioactive implants, 3D bioprinting and tissue engineering mean that implants containing stem cells are surviving longer and undergoing lower levels of immune rejection and human trials are becoming more feasible. But even with the increase in the number of studies investigating the benefit of stem cell use in humans, large scale randomised control trials and cohort studies with long term follow will be needed to assess the true potential of stem cells before they can truly be considered as a viable therapy of the future.

References

1. Reubinoff BE, Pera MF, Fong CY, Trounson A, Bongso A. Embryonic stem cell lines from human blastocysts: somatic differentiation in vitro. *Nat Biotechnol.* Apr 2000;18(4):399-404. doi:10.1038/74447
2. Pittenger MF. Mesenchymal stem cells from adult bone marrow. *Methods Mol Biol.* 2008;449:27-44. doi:10.1007/978-1-60327-169-1_2
3. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell.* Aug 25 2006;126(4):663-76. doi:10.1016/j.cell.2006.07.024
4. Chin MH, Mason MJ, Xie W, et al. Induced pluripotent stem cells and embryonic stem cells are distinguished by gene expression signatures. *Cell Stem Cell.* Jul 2 2009;5(1):111-23. doi:10.1016/j.stem.2009.06.008
5. Bedel A, Beliveau F, Lamrissi-Garcia I, et al. Preventing Pluripotent Cell Teratoma in Regenerative Medicine Applied to Hematology Disorders. *Stem Cells Transl Med.* Feb 2017;6(2):382-393. doi:10.5966/sctm.2016-0201
6. Dominici M, Le Blanc K, Mueller I, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy.* 2006;8(4):315-7. doi:10.1080/14653240600855905

7. Maleki M, Ghanbarvand F, Reza Behvarz M, Ejtemaei M, Ghadirkhomi E. Comparison of mesenchymal stem cell markers in multiple human adult stem cells. *Int J Stem Cells*. Nov 2014;7(2):118-26. doi:10.15283/ijsc.2014.7.2.118
8. Jia Z, Liu Q, Liang Y, et al. Repair of articular cartilage defects with intra-articular injection of autologous rabbit synovial fluid-derived mesenchymal stem cells. *J Transl Med*. May 9 2018;16(1):123. doi:10.1186/s12967-018-1485-8
9. Ullah I, Subbarao RB, Rho GJ. Human mesenchymal stem cells - current trends and future prospective. *Biosci Rep*. Apr 28 2015;35(2)doi:10.1042/BSR20150025
10. Chen L, Tredget EE, Wu PY, Wu Y. Paracrine factors of mesenchymal stem cells recruit macrophages and endothelial lineage cells and enhance wound healing. *PLoS One*. Apr 2 2008;3(4):e1886. doi:10.1371/journal.pone.0001886
11. Kern S, Eichler H, Stoeve J, Kluter H, Bieback K. Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue. *Stem Cells*. May 2006;24(5):1294-301. doi:10.1634/stemcells.2005-0342
12. Ryan JM, Barry FP, Murphy JM, Mahon BP. Mesenchymal stem cells avoid allogeneic rejection. *J Inflamm (Lond)*. Jul 26 2005;2:8. doi:10.1186/1476-9255-2-8
13. Xue L, Xu YB, Xie JL, et al. Effects of human bone marrow mesenchymal stem cells on burn injury healing in a mouse model. *Int J Clin Exp Pathol*. 2013;6(7):1327-36.
14. Atalay S, Coruh A, Deniz K. Stromal vascular fraction improves deep partial thickness burn wound healing. *Burns*. Nov 2014;40(7):1375-83. doi:10.1016/j.burns.2014.01.023
15. Caplan AI. Cell delivery and tissue regeneration. *Journal of Controlled Release*. 1990;11(1-3):157-165. doi:10.1016/0168-3659(90)90129-h
16. Caplan AI. Adult mesenchymal stem cells for tissue engineering versus regenerative medicine. *J Cell Physiol*. Nov 2007;213(2):341-7. doi:10.1002/jcp.21200
17. Caplan AI. Mesenchymal stem cells. *J Orthop Res*. Sep 1991;9(5):641-50. doi:10.1002/jor.1100090504
18. Caplan AI. Review: mesenchymal stem cells: cell-based reconstructive therapy in orthopedics. *Tissue Eng*. Jul-Aug 2005;11(7-8):1198-211. doi:10.1089/ten.2005.11.1198
19. O'Brien FJ. Biomaterials & scaffolds for tissue engineering. *Materials Today*. 2011;14(3):88-95. doi:10.1016/s1369-7021(11)70058-x
20. Vuola J, Goransson H, Bohling T, Asko-Seljavaara S. Bone marrow induced osteogenesis in hydroxyapatite and calcium carbonate implants. *Biomaterials*. Sep 1996;17(18):1761-6. doi:10.1016/0142-9612(95)00351-7
21. Gao P, Zhang H, Liu Y, et al. Beta-tricalcium phosphate granules improve osteogenesis in vitro and establish innovative osteo-regenerators for bone tissue engineering in vivo. *Sci Rep*. Mar 22 2016;6:23367. doi:10.1038/srep23367
22. Padaszynski P, Aleksander-Konert E, Zajdel A, et al. Changes in expression of cartilaginous genes during chondrogenesis of Wharton's jelly mesenchymal stem cells on three-dimensional biodegradable poly(L-lactide-co-glycolide) scaffolds. *Cell Mol Biol Lett*. 2016;21:14. doi:10.1186/s11658-016-0012-2
23. Richardson SM, Curran JM, Chen R, et al. The differentiation of bone marrow mesenchymal stem cells into chondrocyte-like cells on poly-L-lactic acid (PLLA) scaffolds. *Biomaterials*. 2006;27(22):4069-4078. doi:10.1016/j.biomaterials.2006.03.017
24. Diomedea F, Gugliandolo A, Cardelli P, et al. Three-dimensional printed PLA scaffold and human gingival stem cell-derived extracellular vesicles: a new tool for bone defect repair. *Stem Cell Res Ther*. Apr 13 2018;9(1):104. doi:10.1186/s13287-018-0850-0
25. Webb WR, Dale TP, Lomas AJ, et al. The application of poly(3-hydroxybutyrate-co-3-hydroxyhexanoate) scaffolds for tendon repair in the rat model. *Biomaterials*. 2013;34(28):6683-6694. doi:10.1016/j.biomaterials.2013.05.041
26. Williams DF. On the mechanisms of biocompatibility. *Biomaterials*. Jul 2008;29(20):2941-53. doi:10.1016/j.biomaterials.2008.04.023

27. Williams DF. Specifications for Innovative, Enabling Biomaterials Based on the Principles of Biocompatibility Mechanisms. *Front Bioeng Biotechnol.* 2019;7:255. doi:10.3389/fbioe.2019.00255
28. Moshkbouymatin N. *Identification and characterisation of tracheal cartilage derived stem cells for airway tissue engineering.* thesis. Swansea University; 2019.
29. Docheva D, Padula D, Popov C, Mutschler W, Clausen-Schaumann H, Schieker M. Researching into the cellular shape, volume and elasticity of mesenchymal stem cells, osteoblasts and osteosarcoma cells by atomic force microscopy. *Journal of Cellular and Molecular Medicine.* 2008;12(2):537-552. doi:10.1111/j.1582-4934.2007.00138.x
30. Schreivogel S, Kuchibhotla V, Knaus P, Duda GN, Petersen A. Load-induced osteogenic differentiation of mesenchymal stromal cells is caused by mechano-regulated autocrine signaling. *Journal of Tissue Engineering and Regenerative Medicine.* 2019;13(11):1992-2008. doi:10.1002/term.2948
31. Alves da Silva ML, Martins A, Costa-Pinto AR, et al. Chondrogenic differentiation of human bone marrow mesenchymal stem cells in chitosan-based scaffolds using a flow-perfusion bioreactor. *Journal of Tissue Engineering and Regenerative Medicine.* 2011;5(9):722-732. doi:10.1002/term.372
32. Gohin S, Javaheri B, Hopkinson M, Pitsillides AA, Arnett TR, Chenu C. Applied mechanical loading to mouse hindlimb acutely increases skeletal perfusion and chronically enhanced vascular porosity. *Journal of Applied Physiology.* 2020;128(4):838-846. doi:10.1152/jappphysiol.00416.2019
33. Gordon KJ, Blobe GC. Role of transforming growth factor- β superfamily signaling pathways in human disease. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease.* 2008;1782(4):197-228. doi:10.1016/j.bbadis.2008.01.006
34. Grose R, Dickson C. Fibroblast growth factor signaling in tumorigenesis. *Cytokine & Growth Factor Reviews.* 2005;16(2):179-186. doi:10.1016/j.cytogfr.2005.01.003
35. Dexheimer V, Gabler J, Bomans K, Sims T, Omlor G, Richter W. Differential expression of TGF- β superfamily members and role of Smad1/5/9-signalling in chondral versus endochondral chondrocyte differentiation. *Scientific Reports.* 2016;6(1)doi:10.1038/srep36655
36. Lamplot JD, Qin J, Nan G, et al. BMP9 signaling in stem cell differentiation and osteogenesis. *Am J Stem Cells.* 2013;2(1):1-21.
37. Morgan B, Bauza-Mayol G, Gardner O, et al. BMP9 is a potent chondrogenic and morphogenic factor for articular cartilage-derived chondroprogenitors. *Stem Cells and Development.* 2020;doi:10.1089/scd.2019.0209
38. Ma T, Yang S-T, Kniss DA. Oxygen Tension Influences Proliferation and Differentiation in a Tissue-Engineered Model of Placental Trophoblast-Like Cells. *Tissue Engineering.* 2001;7(5):495-506. doi:10.1089/107632701753213129
39. Mas-Bargues C, Sanz-Ros J, Román-Domínguez A, et al. Relevance of Oxygen Concentration in Stem Cell Culture for Regenerative Medicine. *International Journal of Molecular Sciences.* 2019;20(5)doi:10.3390/ijms20051195
40. Dale TP, Mazher S, Webb WR, et al. Tenogenic Differentiation of Human Embryonic Stem Cells. *Tissue Engineering Part A.* 2018;24(5-6):361-368. doi:10.1089/ten.tea.2017.0017
41. Gómez-Leduc T, Desancé M, Hervieu M, et al. Hypoxia Is a Critical Parameter for Chondrogenic Differentiation of Human Umbilical Cord Blood Mesenchymal Stem Cells in Type I/III Collagen Sponges. *International Journal of Molecular Sciences.* 2017;18(9)doi:10.3390/ijms18091933
42. Rosenthal LHS, Benninger MS, Deeb RH. Vocal fold immobility: A longitudinal analysis of etiology over 20 years. *Laryngoscope.* 2007;117(10):1864-70. doi:10.1097/MLG.0b013e3180de4d49
43. Kazuno A, Maki D, Yamato I, et al. Regeneration of Transected Recurrent Laryngeal Nerve Using Hybrid-Transplantation of Skeletal Muscle-Derived Stem Cells and Bioabsorbable Scaffold. *J Clin Med.* Sep 12 2018;7(9)doi:10.3390/jcm7090276
44. Ahmad I, Akhtar MS. Use of Vein Conduit and Isolated Nerve Graft in Peripheral Nerve Repair: A Comparative Study. *Plast Surg Int.* 2014;2014:1-7. doi:10.1155/2014/587968
45. Li Y, Yu Z, Men Y, Chen X, Wang B. Laminin-chitosan-PLGA conduit co-transplanted with schwann and neural stem cells to repair the injured recurrent laryngeal nerve. *Exp Ther Med.*

2018;16(2):1250–8. doi:10.3892/etm.2018.6343

46. Li Y, Xu W, Cheng LY. Adipose-derived mesenchymal stem cells accelerate nerve regeneration and functional recovery in a rat model of recurrent laryngeal nerve injury. *Neural Regen Res*. 2017;12(9):1544–50. doi: 10.4103/1673-5374.215267.
47. Aziz KM, Yu AK, Chen D, Sekula RF. Management of Cranial Nerve Injuries. In: Schmidek and Sweet Operative Neurosurgical Techniques: Indications, Methods, and Results: Sixth Edition. Elsevier Inc.; 2012. p. 2329–38.
48. Moore AM, Wagner IJ, Fox IK. Principles of nerve repair in complex wounds of the upper extremity [Internet]. Vol. 29, Seminars in Plastic Surgery. *Thieme Medical Publishers, Inc*. 2015; p. 40–7. doi:10.1055/s-0035-1544169
49. Guo BF, Dong MM. Application of neural stem cells in tissue-engineered artificial nerve. *Otolaryngol - Head Neck Surg*. 2009;140(2):159–64. doi: 10.1016/j.otohns.2008.10.039
50. Aggarwal SK, Gupta AK, Modi M, Gupta R, Marwaha N. Safety profile of bone marrow mononuclear stem cells in the rehabilitation of patients with posttraumatic facial nerve paralysis—a novel modality (phase one trial). *J Neurol Surg B Skull Base*. Aug 2012;73(4):245–52. doi:10.1055/s-0032-1312716
51. Honings J, Gaissert HA, Verhagen AF, et al. Undertreatment of tracheal carcinoma: multidisciplinary audit of epidemiologic data. *Ann Surg Oncol*. Feb 2009;16(2):246–53. doi:10.1245/s10434-008-0241-3
52. Qiu J, Lin W, Zhou ML, Zhou SH, Wang QY, Bao YY. Primary small cell cancer of cervical trachea: a case report and literature review. *Int J Clin Exp Pathol*. 2015;8(6):7488–93.
53. Elliott MJ, De Coppi P, Speggorin S, et al. Stem-cell-based, tissue engineered tracheal replacement in a child: a 2-year follow-up study. *Lancet*. Sep 15 2012;380(9846):994–1000. doi:10.1016/S0140-6736(12)60737-5
54. Kim IG, Park SA, Lee SH, et al. Transplantation of a 3D-printed tracheal graft combined with iPS cell-derived MSCs and chondrocytes. *Sci Rep*. 2020;10(1):4326. doi:10.1038/s41598-020-61405-4
55. Zhong Y, Yang W, Yin Pan Z, et al. In vivo transplantation of stem cells with a genipin linked scaffold for tracheal construction. *J Biomater Appl*. 2019;34(1):47–60. doi:10.1177/0885328219839193
56. Olthoff A, Mrugalla S, Laskawi R, et al. Assessment of irregular voices after total and laser surgical partial laryngectomy. *Arch Otolaryngol Head Neck Surg*. Sep 2003;129(9):994–9. doi:10.1001/archotol.129.9.994
57. Leydon C, Selekman JA, Palecek S, Thibeault SL. Human embryonic stem cell-derived epithelial cells in a novel in vitro model of vocal mucosa. *Tissue Eng Part A*. Oct 2013;19(19-20):2233–41. doi:10.1089/ten.TEA.2012.0744
58. Lungova V, Chen X, Wang Z, Kendziorski C, Thibeault SL. Human induced pluripotent stem cell-derived vocal fold mucosa mimics development and responses to smoke exposure. *Nat Commun*. 2019;10(1). doi: 10.1038/s41467-019-12069-w
59. Wingstrand VL, Gronhoj Larsen C, Jensen DH, et al. Mesenchymal Stem Cell Therapy for the Treatment of Vocal Fold Scarring: A Systematic Review of Preclinical Studies. *PLoS One*. 2016;11(9):e0162349. doi:10.1371/journal.pone.0162349
60. Hiwatashi N, Bing R, Kraja I, Branski RC. Mesenchymal stem cells have antifibrotic effects on transforming growth factor- β 1-stimulated vocal fold fibroblasts. *Laryngoscope*. 2017;127(1):E35–41. doi: 10.1002/lary.26121
61. Nagubothu SR, Sugars R V., Tudzarovski N, et al. Mesenchymal stromal cells modulate tissue repair responses within the injured vocal fold. *Laryngoscope*. 2020;130(1):E21–9. doi: 10.1002/lary.27885
62. Mattei A, Magalon J, Bertrand B, et al. Autologous adipose-derived stromal vascular fraction and scarred vocal folds: first clinical case report. *Stem Cell Res Ther*. Jul 27 2018;9(1):202. doi:10.1186/s13287-018-0842-0

63. Wood FM, Kolybaba ML, Allen P. The use of cultured epithelial autograft in the treatment of major burn wounds: eleven years of clinical experience. *Burns*. Aug 2006;32(5):538-44. doi:10.1016/j.burns.2006.02.025
64. Tiwari VK. Burn wound: How it differs from other wounds? *Indian J Plast Surg*. May 2012;45(2):364-73. doi:10.4103/0970-0358.101319
65. Akita S, Hayashida K, Takaki S, Kawakami Y, Oyama T, Ohjimi H. The neck burn scar contracture: a concept of effective treatment. *Burns Trauma*. 2017;5:22. doi:10.1186/s41038-017-0086-8
66. Akita S, Yoshimoto H, Ohtsuru A, Hirano A, Yamashita S. Autologous adipose-derived regenerative cells are effective for chronic intractable radiation injuries. *Radiat Prot Dosimetry*. Oct 2012;151(4):656-60. doi:10.1093/rpd/ncs176
67. Jan SN, Bashir MM, Khan FA, et al. Unfiltered Nanofat Injections Rejuvenate Postburn Scars of Face. *Ann Plast Surg*. 2019;82(1):28–33. doi: 10.1097/SAP.0000000000001631
68. Yates CC, Rodrigues M, Nuschke A, et al. Multipotent stromal cells/mesenchymal stem cells and fibroblasts combine to minimize skin hypertrophic scarring. *Stem Cell Res Ther*. Sep 5 2017;8(1):193. doi:10.1186/s13287-017-0644-9
69. Lisa A, Summo V, Bandi V, et al. Autologous Fat Grafting in the Treatment of Painful Postsurgical Scar of the Oral Mucosa. *Case Rep Med*. 2015;2015:842854. doi: 10.1155/2015/842854.
70. Abou Eitta RS, Ismail AA, Abdelmaksoud RA, Ghezlan NA, Mehanna RA. Evaluation of autologous adipose-derived stem cells vs. fractional carbon dioxide laser in the treatment of post acne scars: a split-face study. *Int J Dermatol*. 2019;58(10):1212–22. doi: 10.1111/ijd.14567
71. Shpichka A, Butnaru D, Bezrukov EA, et al. Skin tissue regeneration for burn injury. *Stem Cell Res Ther*. Mar 15 2019;10(1):94. doi:10.1186/s13287-019-1203-3
72. Hirsch T, Rothoefl T, Teig N, et al. Regeneration of the entire human epidermis using transgenic stem cells. *Nature*. Nov 16 2017;551(7680):327-332. doi:10.1038/nature24487
73. Shin SC, Seo Y, Park HY, et al. Regenerative potential of tonsil mesenchymal stem cells on surgical cutaneous defect. *Cell Death Dis*. 2018;9(2). doi: 10.1038/s41419-017-0248-4.
74. Lassig AAD, Lindgren BR, Itabiyi R, Joseph AM, Gupta K. Excessive inflammation portends complications: Wound cytokines and head and neck surgery outcomes. *Laryngoscope*. 2019;129(7):E238–46. doi: 10.1002/lary.27796.