Author (year)	Positive Outcomes	Negative Outcomes	Strengths	Limitations	Suggested study improvements
Liao et al. (2019) (19)	<ul> <li>Auricle stiffness similar to human ear</li> <li>Fusion of diced cartilage created sufficient pinna strength and flexibility</li> <li>Fusion of main pinna structures at 4 months</li> <li>No complications (haematoma, seroma or infection) at implantation site</li> </ul>	<ul> <li>Synthetic scaffold (polyamide) induced a cystic-like reaction subcutaneously</li> <li>Not all structures fused by 4 months</li> <li>Chondrocytes harvested from healthy tissue with donor site morbidity</li> </ul>	<ul> <li>Prospective study</li> <li>Used compression plates for biomechanical strength testing of pinna</li> <li>Histological staining used to assess presence of chondrocytes, matrix and type II collagen</li> <li>Ethical approval</li> </ul>	<ul> <li>Animal study</li> <li>10 subjects</li> <li>12 week end point</li> <li>One scaffold material tested</li> <li>Neocartilage weight used as success marker not considering possible calcification or bone formation</li> <li>Appearance of pinna not assessed (primary outcomes presence of chondrocytes, matrix and strength)</li> <li>Consider difference in pressure exerted on this subcutaneous dorsal pinna vs tight human pinna skin and shape/size changes overtime</li> <li>No testing for cartilage calcification or bone formation</li> </ul>	<ul> <li>Larger numbers with longer follow up</li> <li>Consideration of aesthetics</li> <li>Human application</li> <li>Implant removal and QPCR analysis for bone markers along with alizarin staining to rule out calcification</li> </ul>
Zhou et al. (2018) (22)	<ul> <li>Successfully implanted tissue engineered ear into 5 children</li> <li>Individualised shape based on unaffected ear</li> <li>Shape of bioengineered pinna pre-implantation (12 weeks) showed &gt;90% similarity to the original bioscaffold on laser scanning</li> <li>Pinna contours (helix, triangular fossa, anti-helix and cavum conchae) defined by 9 months</li> <li>Majority of scaffold PGA fibres degraded during 3 months in- vitro prior to implantation, minimising host response</li> <li>PCL core in scaffold significantly improved strength of reconstructed pinna in vitro 4- fold</li> </ul>	<ul> <li>Reconstructed ear stiff and inflexible 12 months post- operatively (note improved by 24 months with further PCL degradation)</li> <li>One of the five subjects lacked evidence of cartilage formation 6 months post-operatively</li> <li>Some surgical implantation methods predispose to graft extrusion</li> </ul>	<ul> <li>Prospective human study</li> <li>Clear primary outcomes of shape, size and cranio-auriculo angle</li> <li>Detailed step-by-step explanation of the 3 methods used</li> <li>Ethical approval</li> <li>SEM used regularly to analyse adherence of chondrocytes to scaffold</li> <li>Quality control sample of the tissue-engineered cartilage used for assessments</li> <li>In vitro method enables quality assessment prior to implantation</li> <li>Shape analysis using 3D laser scanning prior to implantation</li> <li>Regular post-operative review at 1, 2, 3, 6, 9, 12, 18, 24 and 30 months.</li> <li>Histological and immunohistochemical analysis of neocartilage post-operatively (tragal biopsies taken at revision surgeries)</li> </ul>	<ul> <li>Five subjects</li> <li>Only one case followed up in detail (others to follow)</li> <li>Maximum follow up of 2.5 years (other cases ranged 2-18 months), not allowing for complete degradation of scaffold (takes 2-4 years)</li> <li>Single centre</li> <li>This method only suitable for grade II or III microtia, where microtia cartilage can be obtained</li> <li>Tissue expander required for 3 months with psycho-social impact</li> <li>Split-thickness skin graft from groin required at initial procedure</li> <li>Subsequent flap/scar revision surgeries required at 6 and 18 months</li> </ul>	<ul> <li>More subjects</li> <li>Longer follow up (min. 5 years allowing complete scaffold degradation)</li> <li>Use of more mature neocartilaginous grafts recommended, easing surgical handling</li> <li>Multicentre trial</li> <li>Standardised surgical method of implantation</li> <li>Histological analysis including alizarin to rule out calcification/hypertrophy and bone formation pre-implantation</li> <li>MRI/CT scan to identify possible calcification of implant once in-situ</li> </ul>
Zopf et al. (2018) (57)	• Demonstrates the effect of scaffold microarchitecture on cartilage formation. Greater chondrogenicity using regular spherical micropores vs random pore placement within scaffold.	Not disclosed	<ul> <li>Prospective study</li> <li>Ethical approval</li> <li>Clear methods</li> <li>Primary outcome of chondrogenesis clearly assessed</li> <li>Histological analysis of auricle following removal</li> </ul>	<ul> <li>Animal study</li> <li>4 week end point</li> <li>Study size unknown</li> <li>No testing for cartilage calcification or bone formation</li> <li>No strength testing</li> </ul>	<ul> <li>Longer duration allowing for shape and size changes</li> <li>QPCR analysis for bone markers along with alizarin staining to rule out calcification</li> </ul>

<i>Pomerant</i> <i>seva et al.</i> (2016) (58)	<ul> <li>High-quality neocartilage formed by 12 weeks confirmed by GAG, collagen type II and elastin</li> <li>Superior neocartilage in the group expanded using bFGF- expanded chondrocytes</li> </ul>	<ul> <li>Some resorption of neocartilage before 6 weeks</li> <li>Some shrinkage of neocartilage after 20 weeks</li> <li>Poor elastin formation at 12 weeks in group not expanded</li> </ul>	<ul> <li>Aesthetics considered as secondary outcome considering auricular contours, dimensions and projection</li> <li>Clear description of scaffold design and in vitro methods</li> <li>Histological and immunohistochemical analysis with full thickness punch biopsies</li> <li>Prospective study</li> </ul>	<ul> <li>Consider difference in pressure exerted on this subcutaneous pinna vs tight human pinna skin and shape/size changes overtime</li> <li>Animal study</li> <li>Short follow up (6, 12, 20 weeks)</li> <li>Punch biopsies used for analysis rather than full size cross sections showing contiguous neocartilage (due to titanium wire)</li> </ul>	<ul> <li>Longer follow up allowing for shape and size changes</li> <li>Implant removal and QPCR analysis for bone markers along with alizarin staining to rule out calcification</li> </ul>
	<ul> <li>Enhanced elastin fibre quality in neocartilage formed from bFGF-expanded chondrocytes</li> <li>Neocartilage quality improved with time from implantation</li> <li>≤ 10% dimension change at 20 weeks</li> <li>No neocartilage resorption from 6-20 weeks</li> </ul>	with bFGF	• Ethical approval	<ul> <li>Scaffold implanted subcutaneously in neck and subject to less pressure than post-auricular placement</li> <li>No testing for cartilage calcification or bone formation</li> </ul>	
Zopf et al. (2015) (59)	<ul> <li>Histology of in vitro constructs showed cartilage-like tissue at end point</li> <li>Patient-specific auricles using CT scanning and CAD</li> </ul>	• Incomplete cartilage fusion at end point (2 months)	<ul> <li>2 institutions</li> <li>Ethical approval</li> <li>Prospective study</li> <li>Clear description of scaffold design and in vitro methods</li> <li>Histological analysis and staining of auricles</li> <li>Scaffolds implanted post- auricularly, more realistic than on animal dorsum</li> </ul>	<ul> <li>Animal study</li> <li>8 week end point</li> <li>Study size unknown</li> <li>Lack of comparison of neocartilage to native cartilage</li> <li>Poor description of in vivo application</li> <li>No testing for cartilage calcification or bone formation</li> </ul>	<ul> <li>Separate studies for in vitro and in vivo application</li> <li>Longer duration allowing for shape and size changes</li> <li>Implant removal and QPCR analysis for bone markers along with alizarin staining to rule out calcification</li> </ul>
Bichara et al. (2014) (60)	<ul> <li>Neocartilage identified throughout cross section of construct with even distribution</li> <li>Optimal ovine neocartilage formation following 2 weeks in vitro culture</li> <li>Neocartilage quality improved with increased implantation time</li> <li>Ovine elastin detected at 12 weeks in vivo</li> <li>No scaffold extrusion, localised swelling or erythema</li> </ul>	<ul> <li>Neocartilage quality reduced with increased in vitro culture duration</li> <li>Ovine neocartilage showed significantly reduced GAG compared to native cartilage</li> <li>Some shrinkage of construct at 6 weeks in vitro</li> </ul>	<ul> <li>Clear description of scaffold design and in vitro methods</li> <li>Histological analysis and staining of auricles</li> <li>Prospective study</li> <li>Ethical approval</li> </ul>	<ul> <li>Animal study</li> <li>12 week end point</li> <li>Scaffold implanted subcutaneously in neck and subject to less pressure than post-auricular placement</li> <li>No testing for cartilage calcification or bone formation</li> </ul>	<ul> <li>Longer follow up allowing for shape and size changes</li> <li>Implant removal and QPCR analysis for bone markers along with alizarin staining to rule out calcification</li> </ul>
Sterodima s et al. (2013) (61)	<ul> <li>Ears maintained shape and flexibility</li> <li>Histological analysis showed evidence of cartilage formation, type II collagen and matrix</li> <li>No extrusion or infection of implants</li> </ul>	• All showed reduction in both pinna height and width after 8 weeks	<ul> <li>Animals treated as per international guidance</li> <li>Prospective</li> <li>Clear methods</li> <li>Primary outcomes of shape, size and histology addressed</li> </ul>	<ul> <li>Animal study</li> <li>6 subjects</li> <li>8 week end point</li> <li>Pinna flexibility measured subjectively using forceps</li> <li>Aesthetics not considered</li> </ul>	<ul> <li>Larger sample</li> <li>Longer duration allowing for shape and size changes</li> <li>QPCR analysis following implant removal for bone markers along with alizarin staining to rule out calcification</li> </ul>

Yanaga et al. (2009) (21)	<ul> <li>First successful human implantation of regenerated cartilage tissue</li> <li>Neocartilage showed adequate strength and elasticity for auricle reconstruction</li> <li>No evidence of neocartilage absorption over 2-5 year follow up</li> <li>Immunohistochemistry showed evidence of type II collagen</li> </ul>	Variable surgical techniques for shaping cartilage	<ul> <li>Ethical approval</li> <li>Prospective</li> <li>Clear methods</li> <li>Microtia cartilage used hence minimal donor site morbidity</li> <li>Reduced chance of rejection given autologous cartilage</li> </ul>	<ul> <li>4 subjects</li> <li>50% of subjects underweight, potential impact on wound healing</li> <li>2-5 years follow up not allowing fully for reabsorption</li> <li>Neocartilage required sculpting by hand therefore range of techniques and outcomes</li> <li>Two-stage implementation hence multiple surgeries</li> </ul>	<ul> <li>Larger sample of patients</li> <li>One surgeon for all cartilage reconstruction surgeries</li> <li>QPCR analysis for bone markers along with alizarin staining to rule out calcification as quality control pre-implantation</li> <li>MRI/CT scan to identify possible calcification of implant once in-situ</li> </ul>
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Table III: Critical Analysis of Literature

Abbreviations: QPCR quantitative polymerase chain reaction, PGA polyglycolic acid; PCL polycaprolactone; SEM scanning electron microscopy; 3D 3 dimensional; MRI magnetic resonance

imaging; CT computed tomography; GAG glycosaminoglycan; bFGF (FGF-2) basic fibroblast growth factor; CAD computer aided design;