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Arbuscular mycorrhizal associations in plant nutrition and health

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Abstract

Plants and arbuscular mycorrhizal fungi have co-evolved over a period of at least 450 million years. This fungal-plant association involves the transfer of carbon to the obligate biotropic fungus, in return for a wide range of beneficial functions. Although this is usually a mutualistic relationship, it can become parasitic to the plant under adverse conditions. Here, the research examining mechanisms by which mycorrhizal associations improve plant fitness is reviewed. Although there is strong evidence that a number of beneficial functions are performed by mycorrhizae, the mechanisms behind these are often not clear. There are numerous factors which influence these mechanisms and their outcomes, one or more of which can be affecting the association simultaneously. The knowledge we have on arbusular mycorrhizal associations with plants could be applied to various land management practices in order to improve soil degradation brought about by anthropogenic activities. These include erosion, drought, nutrient stress and salinization, and are often a result of poor land management. In order to use mycorrhizal fungi as a biomanagement tool, more research is required, particularly in mature field communities over long timescales. There is a need to invest in the development of sustainable agroecological management methods and to design future policy and legislation that encourages large organizations to incorporate more sustainable practices whilst protecting small-scale farmers.

Keywords: Arbuscular, Mycorrhizal, Fungi, Agroecology, Agriculture, Plant nutrition

Review Methodology: The following databases were searched for research articles and review papers: ISI Web of Knowledge, Google Scholar and CAB Abstracts. Topic-specific search terms were used in searches. References cited in the articles obtained by this method were used to check for additional relevant material.

Introduction

One gram of agricultural soil can contain millions of beneficial microorganisms, which improve soil fertility, including bacteria, algae and fungi [1]. In 1981, Jenkinson and Ladd [1] made a conservative estimate that all soil microorganisms constitute a biomass of 500 kg of C per hectare. A more recent study [2] found that fungi account for a fresh biomass of 4000 kg per hectare of temperate pasture soil – greater than bacteria and algae combined. Not only are fungi abundant in the soil – they are also hugely diverse. The vast diversity of soil fungi has interested researchers since Fries [3] suggested in 1825 that fungi may be as speciose as insects, thereby suggesting a figure of over 140 000 species. A generally accepted estimate of 1.5 million species was made by Hawksworth [4], although other studies have suggested that this may be a vast underestimate, with values of up to 9.9 million being given [5].

Fungi established a symbiotic relationship with the root organs in plants of nearly all terrestrial plant ecosystems worldwide [6, 7], and involve up to 80% of all plant families and approximately 150 fungal species [8]. Of the six groups of mycorrhizal fungi – arbuscular, arbutoid, ecto, ericoid, monoptropoid and orchid [6, 9] – arbusuclar mycorrhizal fungi (AMF)–plant interactions are the most common [7, 10, 11] and these are the most prevalent soil microorganisms in natural and agricultural soils [12]. This interaction is thought to date back at least 450 million years, over which time AMF have become obligate biotrophs as they have lost the ability to capture carbon without associating with a plant host [10, 11, 13, 14].

In order to form associations between the soil and the internal structure of the host species, AMF use hyphae branching threadlike filaments, which make up the mycelium - to proliferate throughout the upper soil horizons and link plants [15-17]. During symbiotic association with a host plant, nutrients are exchanged from fungus to plant in branched, tree-like dichotomous structures formed within plant root cortex cells, called arbuscules [15, 17–20]. These structures transfer nutrients in exchange for carbon through a bidirectional mutualism [10, 21-23], where 5-10% of the host carbon is extracted by AMF [24], thus providing a benefit of host association for the fungus [16]. In return AMF can provide numerous beneficial functions for the host, some examples of which being increased nutrient acquisition [21, 25], improved water relations [26–29], protection from pathogens [30] and sequestration of heavy meals [31, 32], amongst many others. However, it is unclear what factors determine either the relative importance of each function to the plant or which of the aforementioned functions AMF is able to provide in any given situation [33, 34].

Although there is evidence for some host or AMF specificity in AMF-plant relationships, this is not always the case [22, 35–39]. Despite such associations usually being mutualistic (beneficial to both), there is evidence that it can be commensalistic (neither favourable nor detrimental to the two individuals), ammenalistic (one species is inhibited whilst the other is not affected) or even parasitic (advantageous to one individual while having a negative effect on the other [39, 40]. For example, Campos-Soriano [41] found that AMF may have evolved the capacity to evade plant defence mechanisms under conditions where plants are not benefiting from an association, whilst keeping the same functionality.

The mechanisms behind the potentially beneficial functions of AMF--plant associations for plant health and nutrition are discussed below. The degree to which the current literature provides a comprehensive understanding of these processes and the factors which affect them is reviewed. Moreover, the importance of each function in terms of land management is debated. Finally, the implications of these findings with respect to future research and land management are argued.

The Common Mycelial Network and Implications for Plant Community Structure

Biodiversity insures ecosystems against declines in productivity by retaining or increasing species diversity – the greater the variety within a community, the more chance there is that the community will continue to function even if some species can no longer survive in the environment [42]. Species diversity can provide important genetic resources, particularly in environments, which exhibit high genetic diversity, such as semi-natural grasslands [43, 44].

Plant community structure can affect diversity of AMF communities [45, 46]. However, mycorrhizal fungi can also alter plant competition and therefore community structure through a 'common mycelial network' of hyphae linking many plants in one community [35, 47-50]. This concept has been described as the 'wood-wide web', where nutrients can flow between parts of the fungi, and potentially between plants [7, 51]. As a result, plant-plant competition for nutrients may be mediated, at least to a degree, through improved nutrient transfer via the common mycelial network [52-55]. Therefore, microbial soil communities have been described as a driver of plant community dynamics [10], where it is a key mechanism for linking biodiversity and ecosystem functioning and may increase plant biodiversity [56, 57]. However, the degree to which a CMN is beneficial to a host plant is speciesdependant [45, 48, 55], and this network may allow for 'cheater' species to obtain benefits of the common mycelial network without investing significant amounts of carbon [54, 58].

Soil Erosion

Land degradation is recognized as one of the most important global environmental issues, particularly in arid and semi-arid regions. This degradation is a result of numerous climatic and anthropogenic factors, including erosion, drought, nutrient stress and salinization, and often as a result of poor land management [15, 59, 60]. The loss of agricultural productivity due to soil erosion costs the UK €9.99 million annually alone [61]. The network of mycorrhizal hyphae can improve soil stability by binding it through 'sticky' secretions of glomalin, a proteinaceous substance [62-64], creating an entanglement of microaggregates, which leads to macroaggregate formation [59]. This creates a macroporous soil structure which allows water and air to penetrate and reduces erosion [65-67]. As a result, AMF are thought to be the most important factor affecting soil aggregation [40, 62] and are crucial for soil conservation [68, 69].

The complex network of hyphae produced by AMF can equate to up to 30 m of hyphae per 1 g of soil [70, 71], making a significant contribution to the total fungal biomass in soil [72]. AMF hyphae act as an extension of the plant's own root structure, taking over the role of plant root hairs and creating a more branched root system [73, 74]. These fungal hyphae positively influence ecosystem services associated with the below-ground structure, functioning and carbon sequestration, where a high below-ground biomass results in higher ecosystem stability [75]. Numerous studies have shown that a greater abundance of plant roots and mycorrhizae results in higher carbon sequestration [71, 76, 77]. This can mitigate negative effects of climate change from CO_2 emissions [78, 79].

However, a greater understanding of the processes underlying C sequestration is required in order to understand its potential on a global scale. Then, long-term effects of AMF on carbon storage can be modelled [80].

AMF can be significantly reduced – or lost altogether – under conditions of land degradation. This could be through changes in vegetation composition (due to deforestation, agriculture or revegetation) or through agricultural practices such as tillage reducing the inoculum potential [45, 46, 81, 82]. The abundance and diversity of AMF propagules will decrease over time in degraded soils, where plant hosts rely on being colonized by AMF with long-surviving spores [83]. However, the AMF abundance and diversity can be rapidly restored in these soils through transplanting seedlings already colonized by AMF and managed revegetation [60]. The recovery of these AMF communities in highly degraded or desertified ecosystems is essential to successful restoration.

Nutrient Cycling

As a global ecosystem service, the benefits associated with nutrient cycling were valued at US\$2.3 trillion in 1997 [84], although a revised version of this study suggests that this may be a gross underestimation [85]. Agricultural management practices often include significant additions of fertilizers, herbicides and pesticides, which have been shown to reduce mycorrhizal functioning [86-92]. Although studies estimating phosphate reserves vary widely [93] the some estimates suggest that our global phosphate resources could be exhausted within the next 100 years [94]. A review by Berruti et al. [95] found that AMF could be used as a biomanagement tool, where crops inoculated with AMF required 80% less phosphate fertilizer to produce the same yield. Tawaraya et al. [96] also found that the use of AMF combined with lower phosphate application was significantly cheaper per hectare than traditional phosphate fertilizer applications, and therefore is an economically viable option.

The majority of research investigating mycorrhizal fungi has focused on their ability to improve nutrient uptake, particularly of phosphorus [21]. This is because the enhanced availability of nutrients, chiefly phosphorus and nitrogen, is considered the most important function provided by mycorrhizal fungi [10]. Plants rely on AMF for the capture and transfer of soil nutrients through processes of weathering, dissolution and cycling of mineral nutrients and from mobilization of nutrients from organic substances [97]. Up to 90% of plant P and 20% of plant N can be provided by AMF [98]. However, if the soil-N or soil-P availability rises, plants will allocate less carbon to mycorrhizae as they are less reliant on the fungi for their nutrient acquisition, and mycorrhizal abundance will decline [10, 99].

Phosphorus is a major macronutrient required by plants for numerous processes related to plant growth, seed formation and fruit, vegetable and grain quality [100]. Plant-soluble forms of phosphorus, such as phosphate, are very limited in soil [10, 101], making phosphorus availability the most limiting factor for crop yield in 30–40% of arable soils [102, 103]. The inorganic phosphate that is available is rapidly absorbed by plant roots, resulting in a 'phosphorus depletion zone' surrounding the root. AMF can bypass this zone by proliferating in soil which plant roots are unable to reach – a mechanism, which is particularly important in P-limited soils [10, 16, 98, 104]. Conversely, in conditions where plants are not phosphorus-stressed, colonization and growth of mycorrhizal fungi decreases as the AMF association becomes less beneficial to the plant [105].

Nitrogen is an essential component in chlorophyll and plant proteins and is required for cell division [100]. AMF transfer a significant proportion of N to the plant [106, 107], and have been shown to increase plant utilization of nitrogen [10, 108]. As with phosphorus, mycorrhizae can proliferate decomposing patches of organic matter which plant roots are unable to reach and transfer inorganic N to plant roots via the mycelium in exchange for carbon [10, 109]. Although AMF association mainly involves transfer of ammonium, AMF can also assimilate nitrate and amino acids to the plant [110, 111].

Salinization

It has been estimated that between 45 and 77 million hectares of agricultural land are affected by salinity or sodicity stress globally [112, 113] and salinization of arable land is expected to lead to up to 30% land loss within the next 25 years and 50% by 2050 [114–117]. In saline or sodic soils, poor drainage results in the accumulation of salt on the soil surface, negatively affecting plant growth. Increased concentrations of sodium and chlorine and a reduction in potassium, calcium, phosphate and nitrate result in water and nutritional stress [118].

Although extreme saline or sodic soils have been found to delay spore abundance reduce colonization rate and decrease effectiveness of some mycorrhizal associations with plants [119-121], many AMF species are found naturally in saline soils [122]. A recent meta-analysis of studies analysing the effects of mycorrhizal fungi on salt-stressed plants found an overwhelmingly positive response of salt-stressed plants to AMF inoculation [123]. Total yield, flower count, tiller count, leaf area, root fresh weight, shoot length, fruit fresh weight, leaf weight, leaf count, total dry weight, leaf dry weight, shoot fresh weight, biomass yield, fruit count, plant height, root length, grain yield, stem diameter, fruit dry weight, shoot dry weight, root dry weight, stem weight, grain count, total seed weight and root:shoot ratio were all significantly higher for AMFinoculated plants. Only two variables - shoot:root ratio and shoot growth - showed a significant negative effect.

Numerous mechanisms have been proposed to explain how AMF alleviate salt stress, and many of these

mechanisms may occur simultaneously to improve plant tolerance in saline conditions. AMF can enhance nutrient uptake [124–126] and improve rhizospheric and soil conditions [127]. They can reduce production of plant hormones that slow growth, such as ABA [128], accumulate compatible solutes [129] and produce higher levels of antioxidant enzymes [117, 130]. AMF can increase plant chlorophyll concentration [117, 131–133], increase photosynthetic activity [117, 125, 134] and improve water use efficiency and osmotic adjustment at low water potential [117, 131, 135, 136]. Additionally, changes at the cell level, in membranes and cell wall elasticity, have been recorded [137, 138].

Water Relations

Salinity, drought and increasing temperatures are interlinked as these factors all affect the osmostic component of the plant [139, 140]. They are also the most common abiotic stresses affecting crop plants [29, 141]. Humans intercept approximately 60% of water run-off following precipitation, and use 80% of this for agriculture [142]. There has been recent attention on the potential role of AMF to reverse soil degradation in arid and semi-arid areas through improvement of soil quality and subsequent revegetation of land [59, 143, 144].

One of the main processes by which AMF improve water relations under drought conditions is through the secretion of glomalin, a glycoprotein, which can stabilize soil aggregates and therefore increase water retention [63, 145, 146]. However, mycorrhizal fungi are also able to improve water relations directly through transporting water to the plant via fungal hyphae in areas of soil inaccessible to plant roots [97, 147] subsequently improving stomatal control and reducing transpiration rates [135, 147, 148]. The extensive nature of the hyphal network not only leads to greater proliferation into previously inaccessible patches of soil, but also results in a larger surface area for absorption of water (and nutrients) and greater longevity of absorption [149-151]. There is evidence that mycorrhizal hyphae promote plant root development, which leads to improve water uptake [28, 152]. AMF can stimulate the expression of aquaporins - proteinic channels, which facilitate passive water flow and are responsible for cytosolic osmoregulation and water transport [29, 141, 148, 153]. AMF have been shown to increase plant root hydraulic conductivity and to improve water use efficiency via increased nutrient uptake, resulting in more drought-resistant plants [28, 29, 154-157].

Protection Against Soil and Above-ground Organisms

In the USA, the annual cost to agriculture due to nonindigenous species of plants, animals and microbes

was in excess of US\$138 billion annually [158]. Soil-borne pathogens such as nematodes and pathogenic fungi cause significant damage to plants with a high economic importance, such as agricultural crops [158-161]. In order to reduce the negative effects of plant-pathogen interactions, plants exhibit numerous defence responses, which are brought about by their association with a fungal partner. Cell wall thickening occurs when the plant increases synthesis of chitinases and glucanases [162, 163] and the plant can produce a biochemical response, which can alter root structure and exudate composition [164, 165]. Direct competition with root pathogens for colonization sites and altered soil biota may also reduce the negative effects of pathogens on plants [21, 30, 166, 167]. However, recent research has suggested that competition for colonization sites is not the main mechanism by which AMF inhibits soil-borne pathogens [168]. It is likely that there is a cumulative effect from improvement of plant nutrition and from increased resistance through AMF-induced plant defence responses [30], which drives plant pathogen resistance under AMF innoculation.

Biotic reactions among plants and microorganisms below-ground may be equally - if not more - significant than above-ground reactions in determining the outcome of competition between plant species [40, 169-171]. Pineda et al. [171] suggested that it is now widely accepted that 'plant interactions belowground orchestrate a cascade of events that affects the interactions of plants with organisms that live aboveground, and vice versa'. Above-ground ecosystems have tended to be considered separate from below-ground ecosystems [172], however there has been recent increased interest in the interaction between soil organisms and above-ground organisms. There is evidence to suggest that fungi may trigger an indirect plant defence response against herbivores, and vice versa [173-176] since plant defence response to insect predation is not limited to the roots and can result in accumulation of anti-feedant compounds in shoots [126, 127] and up-regulation of genes associated with plant defence [177, 178]. However, AMF is not entirely selfless in its mechanisms of protection: removal of aboveground biomass by herbivores can suppress AMF by altering the plant carbon allocation due to preferential allocation of carbon to other plant parts rather than plant roots [179].

The effects of mycorrhizal colonization vary depending on the organism attacking the plant. For example, a meta-analysis of insect herbivores found that chewing insects and leaf miners were not significantly affected by mycorrhizal colonization, whereas mycorrhizae positively affected sucking insects and negatively affected gall-forming insects [180]. Pozo *et al.* [177] suggested that generalist insects are more strongly affected by plant defence responses than specialists, which can evade these mechanisms. When there is a positive outcome, effects have been linked to improved plant palatability, whereas negative effects are associated with reduced palatability or plant defence responses [181]. However, a recent meta-analysis found that studies need to consider the three-way interactions between plants, microbes and insects. For instance, insects may affect the abundance, susceptibility or accessibility of plants to microbial symbionts and the plant-microbe interactions. Similarly, plants may alter insect-microbe interactions through alterations in food quality for herbivore or susceptibility of insects to plant pathogens [174].

As a result of fungi-induced plant protection, Gianinazzi and Gianinazzi-Pearson [182] described mycorrhizal fungi as 'health insurance' for plants. As a result, mycorrhizal fungi could be used as a biocontrol agent to reduce negative effects of soil and above-ground organisms on plants [174, 183–186]. A review of current literature found that mycorrhiza-induced biocontrol was enhanced under conditions of abiotic stress such as drought, nutrient limitation and salinity, therefore mycorrhizal associations may become more important over time as biotic and abiotic stresses on plants are expected to increase [187]. However, their actual use as a biological control agent is still limited as success varies depending on the AMF isolate, pathogen, plant and environmental conditions [188, 189]. More research is required to develop a comprehensive understanding of the potential role of AMF.

Remediation of Heavy-metal Contaminated Soils

In natural conditions, heavy metals are found at low concentrations in rock and soils, posing no significant environmental risk [190]. Many heavy metals are required by plants in small concentrations in order to act as enzyme cofactors or to maintain a functional plant metabolism; however, some heavy metals such as cadmium have no known benefit to plants [191–193]. High concentrations of heavy metals can result in reduced plant growth, changes to mineral concentrations in plant tissues, root browning and altered photosynthesis [194]. Heavy metal contamination of soils has increased due to industrial and agricultural practices such as mining, smelting, industrial effluents, manufacturing and processing of goods, and addition of natural and synthesized fertilizers in agriculture [18].

A number of remediation technologies exist to treat contaminated soils, such as excavation and subsequent land fill, thermal treatment, electro reclamation, soil washing, vitrification, acid leaching, evaporation, ion exchange and solvent extraction [31, 32]. However these methods are expensive and inefficient, and have been found to negatively affect numerous soil properties and destroy the majority of organisms within the soil [31, 32, 195]. Bioremediation is suggested as a viable alternative [196, 197], particularly using phytoremediation by plants through phytostabilization (stabilizing pollutants through immobilization) phytodegradation (plant metabolic processes break down pollutants) and phytoextraction (pollutants hyperaccumulate in plant tissues which are then harvested) [31].

AMF are abundant even in highly degraded soils [198]. Under heavy metal stress, AMF associations resulted in less uptake of heavy metals in plant tissues, better growth and internal detoxification of metals [199, 200]. However, Audet and Charest [201] suggested that the remediation mechanisms may depend on the heavy metal concentration in the soil. The production of glomalin, fungal polyphosphates, phytochelatins and metallothioneins by AMF could result in chelation of toxins, reducing the plant-available heavy metals [202-204]. Fungal colonization can reduce plant root access to heavy metals due to fungal sheath cover at the root surface [205], and the large biomass of AMF can dilute the heavy metal concentration [206]. Fungi may reduce transport of heavy metals through immobilization and compartmentalization via absorption into hyphal walls, reducing concentrations in above-ground plant tissues or accumulating in hyphal walls in a non-toxic form [206–209]. They have also been found to sequester heavy metals in plant roots, preventing translocation to shoots [210-213]. The ability to immobilize heavy metals in the fungal mycelium is thought to be the main protection mechanism for plants in contaminated soils [208, 214]. Accumulation of contaminants can also occur through fungal structures such as arbuscules, vesicles and vacuoles, minimizing toxicity in the plant itself [191]. Finally, since AMF leads to enhanced plant nutrition and water availability resulting in an increase in plant yield, AMF may indirectly dilute the effects of heavy metals by promoting plant growth [208, 215].

Increased heavy metal contamination has often been shown to cause a decrease in mycorrhizal species diversity [216], spore abundance, colonization rates and growth of the extraradical mycelium [217]. In some cases AMF has been completely eradicated under conditions of heavy metal pollution [218]. However, mycorrhizal communities are generally able to recover from the initial inhibition as immobilization limits toxicity and changes in community structure leads to more tolerant organisms [219]. Effective use of mycorrhizal fungi in bioremediation requires an understanding of the AMF species present in the soil at a given contaminated site, since AMF will vary in their ecological diversity, functional compatibility with phtoremediation plants and sensitivity to heavy metal contamination [31, 200, 220]. Although numerous underlying mechanisms for improved plant tolerance through AMF associations have been suggested, these are still poorly understood and require further research [221].

Plant Yield and Reproductive Structures

A major indicator of plant nutrition and health is yield, particularly for economically important crop and tree species. However, it may be more useful to examine the effects of a stressor on root:shoot ratio, rather than investigating changes in above- and belowground biomass. Resource allocation to roots has been shown to regulate intensity of formation of mycorrhizal structures and carbon availability to the fungus [99, 222–224]. Conversely, it has been suggested that a decrease in mycorrhizal colonization could lead to a reduction in the amount of carbohydrates allocated to roots and a reduction in the size of the common mycelial network [10, 225]. This reduction would lead to a decrease in the root biomass and thus the root: shoot ratio [226, 227]. Studies have found that plant dependence on mycorrhizal fungi may increase as greater root branching causes more resources to be allocated below-ground to roots and hyphae [73, 74].

Although biomass is important for a number of plant species, the effects on reproductive structures, particularly fruits and seeds, can have a significant effect the horticulture industry, which depends on the formation these structures. A reduction in allocation to reproductive structures can negatively affect plant success over multiple years. However, the effects of AMF association on reproductive structures do not always mirror the effects in nutrition and yield [228]. This is because resource allocation may differ for various plant parts, depending on a multitude of factors. For example, removal of above-ground biomass can cause the plant to preferentially allocate carbon away from the roots to other plant parts, resulting in altered carbon allocation to AMF [179]. Conversely, increased growth of plant reproductive structures results in a greater requirement for resources in order to produce sufficient branches, leaves and roots [229]. Mycorrhizal fungi have been shown to affect economically important plants, for example by improving growth of tomato plants and mineral nutrient content of fruits [230].

Management Implications

Approximately 925 million people globally are suffering from malnutrition [231]. Food security is of particular concern in developing countries, where arid climates and poor land management have led to low yields, nutrient deficiencies, soil toxicity and acidity [232]. In Africa, one of the worst-affected regions, the impacts are substantial: 65% of arable land, 30% of grazing land and 20% of forests are already damaged [233].

Agricultural management must incorporate sustainable practices by respecting natural ecological processes and supporting long-term productivity [234]. Since the first 'green revolution', despite an increased interest in the use of mutually beneficial soil microorganisms in agriculture [235], limited attention has been given to the potential contribution of AMF [236]. Although most agricultural crops associate with AMF, intensive management tends to significantly reduce AMF diversity through practices such as monoculture cropping, tillage and fertilizer addition [237–240], although this is not always the case [241].

Fertilizer use is no longer an appropriate management solution to increase nutrient concentrations as this has become more expensive in recent years and some fertilizers are running out [94, 242]. A recent review found that AMF could be used as a biomanagement tool in order to reduce phosphate fertilizer application by up to 80% [95, 96]. Yield has been known to increase when there is a plant–AMF association in stressed environments, such as nutrient deficiency [95], salinity stress [123] and heavy metal pollution [215]. The successful use of plants in soil restoration depends on mycorrhizal associations [200], and it has been demonstrated that a 'phytomicrobial' approach to soil restoration is an economically viable option [96].

In addition to revegetation of degraded land, there is an increasing need to also improve the soil quality [243, 244]. The multiple benefits associated with mycorrhizal fungi ultimately bring about improvements in soil quality and agricultural productivity in areas experiencing severe biotic and abiotic stress [245]. Bethlenfalvay and Linderman [246] stated that 'the role of AMF may be critical if agriculture is to return to the state where luxury levels of farm inputs of fertilizers, pesticides and/or chemicals are decreased to levels that are still economic, yet do not pollute the environment or pose health risks to consumers or handlers'.

In order to incorporate agroecological management practices such as AMF use on a large scale, numerous issues first need to be addressed. Agricultural policy, mainstream trade and land tenure legislation can also no longer punish smallholder farmers, who are the main practitioners of agroecology. Further investment is required to ensure that new approaches to agroecological management are developed, and future policy and legislation should encourage large organizations to incorporate more sustainable practices [247]. These agricultural practices must also be able to strengthen rural communities, improve livelihood of smallholder farmers, and avoid negative social and cultural impacts such as the loss of land tenure and forced migration [248].

Although there have been attempts to develop global policies and legislation on sustainable use of soils, these have not been entirely successful: policies either led to ineffective 'real-life' results or were never implemented due to insufficient international support [249]. Currently, farmers may use negligent, short-sighted or exploitative management practices, while policies may be poorly planned, discriminatory or simply ineffective [250]. In order for mankind to use AMF as a sustainable biomanagement tool to improve degraded soils and reduce malnutrition, the degree to which resources are invested in practitioner education and legislation is as important – if not more so – than investment in research.

Further Research

Although there is a significant body of research on many of the benefits of AMF for plant nutrition and health, there are limitations with current research when attempting to extrapolate results to real-life conditions. These issues can be separated into four key points:

(i) Species diversity

Plants are often grown in low-diversity mixtures for use in pot experiments [180, 251], whereas plant communities are associated with numerous interacting AMF species simultaneously in the field, and vice versa [38]. Since both plants and AMF can preferentially allocate resources to higher quality partners [50, 99] the outcome of an experiment is likely to be strongly dependant on the plant and mycorrhizal species used. Pot experiments have compared mycorrhizal plants with non-mycorrhizal plants [195], however since ~80% of terrestrial plants are associated with mycorrhizal fungi [8] this is not a true representation of natural conditions. Field-based experiments control AMF in this way by either using fungicide treatment in non-AMF plots, which rarely leads to a true 'non-AMF' treatment, or by comparing natural plots to those where AMF has been added [252]. These variances in experimental setup represent a confounding factor for analysis of treatment differences.

(ii) Scale of experiment

While small-scale pot experiments are useful when determining specific interactions of mycorrhizal fungi with a number of biotic and abiotic factors, the outcome of these experiments could be very different in more complex systems [34], for example at the community level in situ. Pot experiments tend to use juvenile plants; however the benefits of mycorrhizal colonization differs depending on the age of plant hosts, where young hosts may receive stronger positive or negative effects from AMF associations compared with species in mature ecosystems [53, 253]. The issues with trying to replicate field conditions in a pot experiment are not limited to issues with plants. For example, since an insect herbivore is rarely selected due to a known preference for a given plant species and mycorrhizal fungi additions, it may not be an interaction seen under natural conditions [180], therefore studies are increasingly placed in a community context [174].

In field experiments, many factors such as changing precipitation, irradiation, temperature and small scale soil properties can confound results [254]. Although pot experiments allows for numerous factors to be controlled, edge effects such as elevated temperature and obstruction can negatively affect plant growth and alter the behaviour of AMF [255]. Pot size may affect root growth, as a lack of space may lead to roots being very crowded in the soil [255, 256]. Nutrient availability can be limiting in pots, restricting plant growth [255]. The effects of AMF may be underestimated in pot experiments, since colonization can be lower when there is a relatively high root density in a confined pot [252]. One promising approach would be to match fungal species with their environmental conditions, for example by tillage regime, soil type, pH or host diversity [80]. Finally, although individual experiments are useful, there is a need for 'big data' research involving the collation

of large quantities fine-scale field data in order to understand global soil quality [257].

(iii) Duration of experiment

The majority of studies on mycorrhizal effects on plants have been conducted over one growing season or less, despite evidence that communities experience phases of vegetation dominance and adapt to environmental changes over timescales significantly longer than this – potentially decades [257–259]. Differences in the duration of the experiment have also been found to lead to variability in response to biotic stressors, such as herbivory [260, 261]. Experiments must consider the temporal variability in abiotic stressors since soil variables such as nutrient concentration [262] and water content [263] vary over time, therefore the duration of the experiment will have a significant impact on the outcome.

(iv) Hierarchies of effects

In order to successfully use AMF to improve degraded soil or increase agricultural productivity, a better understanding of how functional mechanisms differ is necessary [34]. Since numerous variables may interact with one another and affect AMF simultaneously, there is a hierarchy of effects in any given situation depending on the plant stressor(s). Any given variable is controlled by, and controls, a number of factors at any one time, so it would be expected that direct changes in that variable will influence the effects on other variables, and vice versa [264]. Studies can show an overall effect on a given variable, but cannot unequivocally reveal the mechanisms, which cause community-level changes [265]. Therefore Koide [266] stated that 'an understanding of ecologically relevant traits that determine environmentally context-dependent interaction hierarchies is the key to elucidating general principles that structure biological communities'.

Conclusion

AMF receive plant carbon in return for numerous benefits to plant nutrition and health under conditions of stress. However the plant-AMF association is not always mutualistic, and can be parasitic under environmental conditions, which are favourable to the plant. These benefits have implications for a wide range of uses of AMF, particularly as part of agroecological management practices, which aim to restore degraded soils, revegetate land and increase plant yield in a sustainable manner. In order to effectively use these management methods, further research is required, which focuses on studies that can be extrapolated to natural conditions in the field. Although scientific knowledge on the use of AMF in agriculture is useful, translating this knowledge into effective policies has largely failed, particularly at the global scale. If agroecological management is to be successful, advancements need to be made both in our scientific knowledge of biotechnological uses mycorrhizal fungi whilst also educating agricultural practitioners and improving agricultural policy. These policies should

encourage large-scale farmers to manage soil sustainably, whilst allowing the socio-economic status of small-scale farmers to improve.

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References

- Jenkinson DS, Ladd HN. Microbial biomass in soil: measurement and turnover. In: Paul EA, Ladd JN, editors. Soil Biochemistry. 5th ed. Marcel Dekker, Inc., New York, NY, USA; 1981. p. 415–471.
- Pimental D, Stachow U, Takacs DA, Brubaker HW, Dumas AR, Meaney JJ, et al. Conserving biological diversity in agricultural/ forestry systems. BioScience 1992;42:354–62.
- 3. Fries EM. Systema Orbis Vegetabilis. Typographia Academica, Lund, Sweden; 1825.
- Hawksworth DL. The fungal dimension of biodiversity: magnitude, significance, and conservation. Mycological Research 1991;95:641–55.
- Cannon PF. Strategies for rapid assessment of fungal diversity. Biodiversity and Conservation 1997;6:669–80.
- Wang B, Qiu YL. Phylogenetic distribution and evolution of mycorrhizas in land plants. Mycorrhiza 2006;16:299–363.
- Helgason T, Fitter A. Natural selection and the evolutionary ecology of the arbuscular mycorrhizal fungi (*Phylum Glomeromycota*). Journal of Experimental Botany 2009;60:2465–80.
- 8. Fitter AH. Darkness visible, reflections on underground ecology. Journal of Ecology 2005;93:231–43.
- Garg N, Geetanjali, Kaur A. Arbuscular mycorrhizal: nutritional aspects. Archives of Agronomy and Soil Science 2006;52:593–606.
- Smith SE, Read DJ. Mycorrhizal Symbiosis. 3rd ed. Academic Press, London, UK; 2008.
- 11. Redecker D, Kodner R, Graham LE. Glomalean fungi from the Ordovician. Science 2000;289:1920–1.
- Mohammad A, Mittra B. Effects of inoculation with stress-adapted arbuscular mycorrhizal fungus Glomus deserticola on growth of *Solanum melogena L*. and *Sorghum sudanese Staph*. seedlings under salinity and heavy metal stress conditions. Archives of Agronomy and Soil Science 2013;59(2):173–83.
- Requena N, Serrano E, Ocón A, Breuninger M. Plant signals and fungal perception during arbuscular mycorrhizal establishment. Phytochemistry 2007;68:33–40.
- Bonfante P, Genre A. Plants and arbuscular mycorrhizal fungi: an evolutionary developmental perspective. Trends in Plant Science 2008;13:492–8.
- Dodd JC. The role of arbuscular mycorrhizal fungi in agro- and natural ecosystems. Outlook on Agriculture 2000;29:63–70.

- Smith SE, Jakobsen I, Gronlund M, Smith FA. Roles of mycorrhizas in plant phosphorus nutrition: interactions between pathways of phosphorus uptake in arbuscular mycorrhizal roots have important implications for understanding and manipulating plant phosphorus acquisition. Plant Physiology 2011;156:1050–7.
- 17. McNear Jr DH. The rhizosphere roots, soil and everything in between. Nature Education Knowledge 2013;4(3):1
- He X, Nara K. Element biofortification: can mycorrhizas potentially offer a more effective and sustainable pathway to curb human malnutrition? Trends in Plant Science 2007;12:331–3.
- Manchanda G, Garg N. Endomycorrhizal and rhizobial symbiosis: how much do they share? Journal of Plant Interactions 2007;2:79–88.
- Gutjahr C, Parniske M. Cell and developmental biology of Arbuscular Mycorrhiza symbiosis. Annual Review of Cell and Developmental Biology 2013;29:593–617.
- Parniske M. Arbuscular mycorrhiza: the mother of plant root endosymbiosis. Nature Reviews Microbiology 2008;6:763–75.
- 22. Kiers ET, Duhamel M, Beesetty Y, Mensah JA, Franken O, Verbruggen E, *et al.* Reciprocal rewards stabilize cooperation in the mycorrhizal symbiosis. Science 2011;333:880–2.
- Fellbaum CR, Gachomo EW, Beesetty Y, Choudhari S, Strahan GD, Pfeffer PE. Carbon availability triggers fungal nitrogen uptake and transport in arbuscular mycorrhizal symbiosis. Proceedings of the National Academy of Sciences 2011;109(7):2666–71.
- Johnson D, Leake JR, Read DJ. Role of arbuscular mycorrhizal fungi in carbon and nutrient cycling in grassland. In: Gadd GM, editor. Fungi in Biogeochemical Cycles. Cambridge University Press, Cambridge, UK; 2006. p. 129–50.
- Hodge A, Storer K. Arbuscular mycorrhiza and nitrogen: implications for individual plants through to ecosystems. Plant and Soil 2015;386:1–19.
- Zhu XC, Song FB, Liu SQ, Liu TD, Zhou X. Arbuscular mycorrhizae improves photosynthesis and water status of *Zea mays* L. under drought stress. Plant, Soil and Environment 2012;58(4):186–91.
- Birhane E, Sterck FJ, Fetene M, Bongers F, Kuyper TW. Arbuscular mycorrhizal fungi enhance photosynthesis, water use efficiency, and growth of frankincense seedlings under pulsed water availability conditions. Oecologia 2012;169(4):895–904.
- Marulanda A, Barea JM, Azcon R. An indigenous drought tolerant strain of *Glomusintraradices* associated with a native bacterium improves water transport and root development in *Retamasphaerocarpa*. Microbial Ecology 2006;52:670–8.
- 29. Augé RM. Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. Mycorrhiza 2001;11:3–42.
- Cameron DD, Neal AL, van Wees SCM, Ton J. Mycorrhiza-induced resistance: more than the sum of its parts? Trend in Plant Science 2013;18(10):539–45.
- Meier S, Borie F, Bolan N, Cornejo P. Phytoremediation of metal-contaminated soils by arbuscular mycorrhizal fungi. Critical Reviews in Environmental Science and Technology 2012;47(7):741–75.

- Mani D, Kumar C. Biotechnological advances in bioremediation of heavy metals contaminated ecosystems: an overview with special reference to phytoremediation. International Journal of Science and Technology 2014;11:843–72.
- Newsham KK, Fitter AH, Watkinson AR. Multi-functionality and biodiversity in arbuscular mycorrhizas. Trends in Ecology and Evolution 1995;10:407–11.
- Sikes BA. When do arbuscular mycorrhizal fungi protect plant roots from pathogens? Plant Signaling and Behavior 2010b;5(6):763–5.
- Bever JD, Dickie IA, Facelli E, Facelli JM, Klironomos J, Moora M, *et al.* Rooting theories of plant community ecology in microbial interactions. Trends in Ecology and Evolution 2010;25:468–78.
- Bever JD, Richardson SC, Lawrence BM, Holmes J, Watson M. Preferential allocation to beneficial symbiont with spatial structure maintains mycorrhizal mutualism. Ecology Letters 2009;12:13–21.
- Verbruggen E, ElMouden C, Jansa J, Akkermans G, Bucking H, West SA, *et al.* Spatial structure and interspecific cooperation: theory and an empirical test using the mycorrhizal mutualism. American Naturalist 2012a;179:133–46.
- Johnson D, Martin F, Cairney JWG, Anderson IC. The importance of individuals: intraspecific diversity of mycorrhizal plants and fungi in ecosystems. New Phytologist 2012;94:614–25.
- Johnson NC, Graham JH. The continuum concept remains a useful framework for studying mycorrhizal functioning. Plant and Soil 2012;363:411–9.
- Klironomos JN. Variation in plant response to native and exotic arbuscular mycorrhizal fungi. Ecology 2003;84:2292–301.
- Campos-Soriano L, García-Garrido JM, Segundo BS. Activation of basal defense mechanisms of rice plants by *Glomus intraradices* does not affect the arbuscular mycorrhizal symbiosis. New Phytologist 2010;188:597–614.
- Yachi S, Loreau M. Biodiversity and ecosystem productivity in a fluctuating environment: the insurance hypothesis. Proceedings of the National Academy of Sciences of the USA 1999;96(4):1463–8.
- Bullock JM, Aronson J, Newton AC, Pywell RF, Rey-Benayas JM. Restoration of ecosystem services and biodiversity: conflicts and opportunities. Trends in Ecology and Evolution 2011;26:541–9.
- Picó FX, van Groenendael J. Large-scale plant conservation in European semi-natural grasslands: a population genetic perspective. Diversity and Distributions 2007;13:920–6.
- Jeffries P, Barea JM. Arbuscular Mycorrhiza a key component of sustainable plant-soil ecosystems. In: Hock B, editor. Mycota Series, IX. The Mycota, Fungal Associations. Springer-Verlag, Berlin, Germany; 2012. p. 95–113.
- Alguacil MM, Torres MP, Torrecillas E, Díaz G, Roldán A. Plant type differently promote the arbuscular mycorrhizal fungi biodiversity in the rhizosphere after revegetation of a degraded, semiarid land. Soil Biology and Biochemistry 2011;43:167–73.
- Barto KE, Hilker M, Muller F, Mohney F. The fungal fast lane: common mycorrhizal networks extend bioactive zones of allelochemicals in soils. PLoS ONE 2011;6:e27195.

- Lin G, McCormack ML, Guo D. Arbuscular mycorrhizal fungal effects on plant competition and community structure. Journal of Ecology 2015;103(5):1224–32.
- Hodge A, Fitter AH. Microbial mediation of plant competition and community structure. Functional Ecology 2301;27(4):865–75.
- Weremijewicz J, Janos DP. Common mycorrhizal networks amplify size inequality in Andropogon gerardii monocultures. New Phytologist 2013;198:203–13.
- Helgason T, Daniell TJ, Husband R, Fitter AH, Young JPW. Ploughing up the wood-wide web. Nature 1998;394(6692):431.
- Simard SW, Durall DM. Mycorrhizal networks: a review of their extent, function, and importance. Canadian Journal of Botany 2004;82:1140–65.
- Van der Heijden MGA, Horton TR. Socialism in soil? The importance of mycorrhizal fungal networks for facilitation in natural ecosystems. Journal of Ecology 2009;97(6):1139–50.
- Ghoul M, Griffin AS, West SA. Toward an evolutionary definition of cheating. Evolution 2014;68:318–31.
- Walder F, Niemann H, Natarajan M, Lehmann MF, Boller T, Wiemken A. Mycorrhizal networks: common goods of plants shared under unequal terms of trade. Plant Physiology 2012;159:789–97.
- Wagg C, Jansa J, Stadler M, Schmid B, van der Heijden MGA. Mycorrhizal fungal identity and diversity relaxes plant-plant competition. Ecology 2011;92:1303–13.
- van der Heijden MGA, Wiemken A, Sanders IR. Different arbuscular mycorrhizal fungi alter coexistence and resource distribution between co-occurring plant. New Phytologist 2003;157:569–78.
- Sanders IR. Preference, specificity and cheating in the arbuscular mycorrhizal symbiosis. Trends in Plant Science 2003;8:143–5.
- Al-Karaki GN. The Role of mycorrhiza in the reclamation of degraded lands in arid environments. In: Shahid, SA, Taha, FK, Abdelfattah, MA, editors. Developments in Soil Classification, Land Use Planning and Policy Implications: Innovative Thinking of Soil Inventory for Land Use Planning and Management of Land Resources. Springer, Dordrecht, Netherlands; 2013. p. 823–36.
- Dodd JC, Arias I, Koomen I, Hayman DS. The management of populations of vesicular-arbuscular mycorrhizal fungi in acid-infertile soils of a savanna ecosystem. I. The effect of precropping and inoculation with VAM-fungi on plant growth and nutrition in the field. Plant and Soil 1990;122:229–40.
- Görlach B, Landgrebe-Trinkunaite R, Interwies E, Bouzit M, Darmendrail D, Rinaudo JD. Assessing the ecomic impacts of soil degradation. In: Executive Summary Study commissioned by the European Commission. DG Environment, Volume IV, 2004. Study Contract ENV.B.1/ETU/2003/0024. Ecologic: Berlin. p. 1–37.
- Rillig MC, Wright SF, Nichols KA, Schmid WF, Torn MS. The role of arbuscular mycorrhizal fungi and glomalin in soil aggregation: comparing effects of five plant species. Plant and Soil 2002;238:325–33.
- 63. Bedini S, Pellegrino E, Avio L, Pellegrini S, Bazzoffi P, Argese E, *et al.* Changes in soil aggregation and glomalinrelated soil protein content as affected by the arbuscular mycorrhizal fungal species Glomus mosseae and

Glomus intraradices. Soil Biology and Biochemistry 2009;41:1491–6.

- Wu Q, Cao M, Zou Y, He X. Direct and indirect effects of glomalin, mycorrhizal hyphae, and roots on aggregate stability in rhizosphere of trifoliate orange. Scientific Reports 2014;4(5823):1–8.
- Thiet RK, Frey SD, Six J. Do growth yield efficiencies differ between soil microbial communities differing in fungal:bacterial ratios? Reality check and methodological issues. Soil Biology and Biochemistry 2006;38:837–44.
- 66. Schmid T, Meyer J, Oehl F. Integration of mycorrhizal inoculum in high alpine revegetation. In: Feldman F, Kapulnik Y, Baar J, editors. Mycorrhiza Works. Proceedings of the International Symposium Mycorrhiza for Plant Vitality and the Joint Meeting of Working Groups 1–4 of COST Action 870, 3–5 October 2007; Hannover, Germany. Deutsche Phytomedizinische Gesellschaft, Braunschweig, Germany; 2008. p. 278–88.
- 67. Curaqueo G, Barea JM, Acevedo E, Rubio R, Cornejo P, Borie F. Effects of different tillage system on arbuscular mycorrhizal fungal propagules and physical properties in a Mediterranean agroecosystem in central Chile. Soil and Tillage Research 2011;113(1):11–18.
- Kohler J, Caravaca F, Roldan A. An AM fungus and a PGCR intensify the adverse effects of salinity on the stability of the rhizosphere soil aggregates of Lectuca sativa. Soil Biology and Biochemistry 2010;42:429–34.
- Wright SF, Green VS, Cavigelli MA. Glomalin in aggregate size classes from three different farming systems. Soil Tillage Research 2007;8:218–23.
- Cavagnaro TR, Smith FA, Smith SE, Jakobsen I. Functional diversity in arbuscular mycorrhizas: exploitation of soil patches with different phosphate enrichment differs among fungal species. Plant, Cell and Environment; 2005;28:642–50.
- Wilson GWT, Rice CW, Rillig MC, Springer A, Hartnett DC. Soil aggregation and carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal fungi: results from long-term field experiments. Ecology Letters 2009;12:452–61.
- Leake JR, Johnson D, Donnelly D, Muckle G, Boddy L, Read D. Network of power and influence: the role of mycorrhizal mycelium in controlling plant communities and agroecosystem functioning. Canadian Journal of Botany 2004;82:1016–45.
- Orfanoudakis M, Wheeler CT, Hooker JE. Both the arbuscular mycorrhizal fungus *Gigaspora rosea* and Frankia increase root system branching and reduce root hair frequency in *Alnus glutinosa*. Mycorrhiza 2010;20:117–26.
- Wu QS, Liu CY, Zhang DJ, Zou YN, He XE, Wu QH. Mycorrhiza alters the profile of root hairs in trifoliate orange. Mycorrhiza 2016;26(3):237–47.
- Tilman D, Reich PB, Knops JMH. Biodiversity and ecosystem stability in a decade long grassland experiment. Nature 2006;441:629–32.
- Soussana J, Loiseau P, Vuichard N, Ceschia E, Balesdent T, Arrouays D. Carbon cycling and sequestration opportunities in temperate grasslands. Soil Use and Management 2004;20:219–30.
- 77. Kramer S, Marhan S, Haslwimmer H, Ruess L, Kandeler E. Temporal variation in surface and subsoil abundance and function of the soil microbial community in an arable soil. Soil Biology and Biochemistry 2013;61:76–85.

- Treseder KK, Holden SR. Fungal carbon sequestration. Science 2013;339(6127):1528–9.
- Solaiman ZN. Contribution of Arbuscular Mycorrhizal fungi to soil carbon sequestration. In: Solaiman ZM, Abbott LK, Varma A, editors. Mycorrhizal Fungi – Use in Sustainable Agriculture and Land Restoration. Springer, Berlin, Heidelberg, Germany; 2014. p. 287–196.
- Verbruggen E, Veresoglou SD, Anderson IA, Caruso T, Hammer ED, Kohler J, *et al.* Arbuscular mycorrhizal fungi – short-term liability but long-term benefits for soil carbon storage? New Phytologist 2013;197(2):366–8.
- Sälea V, Aguilera P, Laczko A, Mäder P, Berner A, Zihlmann U, et al. Impact of conservation tillage and organic farming on the diversity of arbuscular mycorrhizal fungi. Soil Biology and Biochemistry 2015;84:38–52.
- Mbuthia LW, Acosta-Martínez V, DeBruyn J, Schaeffer S, Tyler D, Odoi E, *et al.* Long term tillage, cover crop, and fertilization effects on microbial community structure, activity: implications for soil quality. Soil Biology and Biochemistry 2015;89:24–34.
- Jeffries P, Craven-Griffiths A, Barea JM, Levy Y, Dodd JC. Application of AMF in the revegetation of desertified Mediterranean ecosystems. In: Gianinazzi S, Schüepp H, Barea JM, Haselwandte K, editors. Mycorrhizal Technology in Agriculture – From Genes to Bioproducts. Birkhauser Verlag, Basel, Switzerland; 2002. p. 151–74.
- Costanza R, d'Arge R, de Groot R, Farber S, Grasso M, Hannon B, *et al.* The value of the world's ecosystem services and natural capital. Nature 1997;387:253–60.
- Costanza R, de Groot R, Sutton P, van der Ploeg S, Anderson SJ, Kubiszewskia I, *et al.* Changes in the global value of ecosystem services. Global Environmental Change 2014;26:152–8.
- Parisabu A, Mohamad RB, Hashim A, Omar D, Morshed M. Effect of herbicide on sporulation and infectivity o vesicular arbuscular mycorrhizal (*Glomus mossae*) symbiosis with Peanut plant. Journal of Animal and Plant Sciences 2013;23(6):1671–8.
- Channabasava A, Lakshman HC, Jorquera MA. Effect of fungicides on association of arbuscular mycorrhiza fungus Rhizophagus fasciculatus and growth of Proso millet (*Panicum miliaceum L.*). Journal of Soil Science and Plant Nutrition 2015;15(1):35–45.
- Chhabra ML, Jalali BL. Impact of pesticides-mycorrhia interaction on growth and development of wheat. Journal of Biopesticides 2013;6(2):129–132.
- Nettles R, Watkins J, Ricks K, Boyer M, Licht M, Atwoods LW, et al. Influence of pesticide seed treatments on rhizosphere fungal and bacterial communities and leaf fungal endophyte communities in maize and soybean. Applied Soil Ecology 2016;102:61–9.
- Zaller JG, Heigl F, Ruess L, Grabmaier A. Glyphosate herbicide affects belowground interactions between earthworms and symbiotic mycorrhizal fungi in a model ecosystem. Scientific Reports 2014;4:5634.
- Makarian H, Poozesh V, Ashgari HR, Nazari M. Interaction effects of arbuscular mycorrhiza fungi and soil applied herbicides on plant growth. Communications in Soil Science and Plant Analysis 2016;47(5):619–29.

- Druille M, Cabello MN, Omacini M, Golluscioa RA. Glyphosate reduces spore viability and root colonization of arbuscular mycorrhizal fungi. Applied Soil Ecology 2013;64:99–103.
- van Kauwenbergh SJ. World Phosphate Rock Reserves and Resources. International Fertilizer Development Center, Muscle Shoals, AL, USA; 2010.
- Cordell D, Drangert JO, White S. The story of phosphorus: global food security and food for thought global environmental change: human and policy dimensions. Global Environmental Change 2009;19:292–305.
- Berruti A, Lumini E, Balestrini R, Bianciotto V. Arbuscular mycorrhizal fungi as natural biofertilizers: let's benefit from past successes. Frontiers in Microbiology 2016;6:1559.
- Tawaraya K, Hirose R, Wagatsuma T. Inoculation of arbuscular mycorrhizal fungi can substantially reduce phosphate fertilizer application to Allium fistulosum L. and achieve marketable yield under field condition. Biology and Fertility of Soils 2012;48(7):839–43.
- 97. Finlay RD. Ecological aspects of mycorrhizal symbiosis: with special emphasis on the functional diversity of interactions involving the extraradical mycelium. Journal of Experimental Botany 2008;59:1115–26.
- Cavagnaro TR, Bender SF, Asghari HR, van der Heijden MGA. The role of arbuscular mycorrhizas in reducing soil nutrient loss. Trends in Plant Science 2015;20(5):283–90.
- 99. Werner GDA, Kiers ET. Partner selection in the mycorrhizal mutualism. New Phytologist 2015;205:1437–42.
- 100. Parker R. Plant and Soil Science: Fundamentals and Applications. Delmar, New York, NY, USA; 2009.
- Bucher M. Functional biology of plant phosphate uptake at root and mycorhiza interfaces. New Phytologist 2007;173:11–26.
- Vance CP, Stone CU, Allan DL. Phosphorus acquisition and use: critical adaptations by plants for securing a non-renewable resource. New Phytologist 2003;157:423–47.
- Barea JM, Ferrol N, Azcón-Aguilar C, Azcón R. Mycorrhizal symbioses. In: White PJ, Hammond JP, editors. The Ecophysiology of Plant-phosphorus Interactions, Vol 7, Plant Ecophysiology Series. Springer, Dordrecht, Netherlands; 2008. p. 143–63.
- Roose T, Fowler AC. A mathematical model for water and nutrient uptake by plant root systems. Journal of Theoretical Biology 2004;228:173–84.
- 105. Verbruggen E, Van der Heijden MGA, Weedon JT, Kowalchuk GA, Röling WFM. Community assembly, species richness and nestedness of arbuscular mycorrhizal fungi in agricultural soils. Molecular Ecology 2012b;21:2341–53.
- He XH, Critchley C, Bledsoe C. Nitrogen transfer within and between plants through common mycorrhizal networks (CMNs). Critical Reviews in Plant Science 2003;22:531–67.
- Leigh J, Hodge A, Fitter AH. Arbuscular mycorrhizal fungi can transfer substantial amounts of nitrogen to their host plant from organic material. New Phytologist 2009;181:199–207.
- 108. Hodge A, Campbell CD, Fitter AH. An arbuscular mycorrhizal fungus accelerates decomposition and acquires nitrogen directly from organic material. Nature 2001;413:297–9.
- Govindarajulu M, Pfeffer PE, Jin HR, Abubaker J, Douds DD, Allen JW, *et al.* Nitrogen transfer in the arbuscular mycorrhizal symbiosis. Nature 2005;435:819–23.

- 110. HaiRu J, Jie L, Jing L, XiaoWei H. Forms of nitrogen uptake, translocation, and transfer via arbuscular mycorrhizal fungi: a review. Science China Life Sciences 2012;55(6):474–82.
- 111. Smith SE, Smith FE. Roles of arbuscular mycorrhizas in plant nutrition and growth: new paradigms from cellular to ecosystem scales. Annual Review of Plant Biology 2011;62:227–50.
- 112. Munns R, Tester M. Mechanisms of salinity tolerance. Annual Review of Plant Biology 2008;59:651–81.
- Sheng M, Tang M, Chan H, Yang B, Zhang F, Huang Y. Influence of arbuscular mycorrhizae on photosynthesis and water status of maize plants under salt stress. Mycorrhiza 2008;18:287–96.
- 114. Rengasamy P. World salinization with emphasis on Australia. Journal of Experimental Botany 2006;57:1017–23.
- Porcel R, Aroca R, Ruiz-Lozano JM. Salinity stress alleviation using arbuscular mycorrhizal fungi. A review. Agronomy for Sustainable Development 2012;32:181–200.
- 116. Kapoor R, Evelin H, Mathur P, Giri B. Arbuscular mycorrhiza: approaches for abiotic stress tolerance in crop plants for sustainable agriculture. In: Tuteja N, Gill SS, editors. Plant Acclimation to Environmental Stress. Springer, New York, NY, USA; 2013. p. 359–401.
- 117. Abdel Latef AA, Chaoxing H. Does the inoculation with *Glomus mosseae* improve salt tolerance in pepper plants? Journal of Plant Growth Regulation 2014;33(3):644–53.
- 118. Bothe H. Arbuscular mycorrhiza and salt tolerance of plants. Symbiosis 2012;58:7–16.
- Juniper S, Abbott LK. Soil salinity delays germination and limits growth of hyphae from propagules of arbuscular mycorrhizal fungi. Mycorrhiza 2006;16:371–9.
- 120. Abdel Latef AA, Chaoxing H. Effect of arbuscular mycorrhizal fungi on growth, mineral nutrition, antioxidant enzymes activity and fruit yield of tomato grown under salinity stress. Scientia Horticulturae 2011;127:228–33.
- 121. Hajiboland R. Role of arbuscular mycorrhiza in amelioration of salinity. In: Ahmad P, Mohamed A, Mahgoub P, editors. Salt Stress in Plants: Signalling, Omics and Adaptations. Springer, New York, NY, USA; 2013. p. 301–54.
- 122. Garcia VI, Mendoza RE. Arbuscular mycorrhizal fungi and plant symbiosis in a saline-sodic soil. Mycorrhiza 2007;17:167–74.
- Jayne B, Quigley M. Influence of arbuscular mycorrhiza on growth and reproductive response of plants under water deficit: a meta-analysis. Mycorrhiza 2014;24(2):109–19.
- Evelin H, Giri B, Kapoor R. Contribution of Glomusintraradices inoculation to nutrient acquisition and mitigat ion of ionic imbalance in NaCI-stressed Trigonellafoenum-graecum. Mycorrhiza 2012;22:203–17.
- Zuccarini P. Mycorrhizal infection ameliorates chlorophyll content and nutrient uptake of lettuce exposed to saline irrigation. Plant, Soil and Environment 2007;53:283–9.
- Al-Karaki GN. Nursery inoculation of tomato with arbuscular mycorrhizal fungi and subsequent performance under irrigation with saline water. Scientia Horticulturae 2006;109:1–7.
- 127. Asghari HR, Marschner P, Smith SE, Smith FA. Growth response of Atriplex nummularia to inoculation with arbuscular mycorrhizal fungi at different salinity levels. Plant and Soil 2005;373:245–56.

- 128. Estrada-Luna AA, Davies FT. Arbuscular mycorrhizal fungi influence water relations, gas exchange, abscisic acid and growth of micropropagated chile ancho pepper (Capsicum annuum) plantlets during acclimatization and post acclimatization. Journal of Plant Physiology 2003;160:1073–83.
- Evelin H, Giri B, Kapoor R. Ultrastructural evidence for AMF mediated salt stress mitigation in Trigonella foenum-graecum. Mycorrhiza 2013;23:71–86.
- Manchanda G, Garg N. Alleviation of salt-induced ionic, osmotic and oxidative stresses in Cajanuscajan nodules by AM inoculation. Plant Biosystems 2011;145:88–97.
- Hajiboland R, Aliasgharzadeh N, Laiegh SF, Poschenreider C. Colonization with arbuscular mycorrhizal fungi improves salinity tolerance of tomato (*Solanum lycopersicum L.*) plants. Plant and Soil 2010;331:313–27.
- 132. Giri B, Mukerji KJ. Mycorrhizal inoculant alleviates salt stress in *Sesbania aegyptiaca* and *Sesbania grandiflora* under field conditions: evidence for reduced and improved magnesium uptake. Mycorrhiza 2004;14:307–12.
- Shekoofeh E, Sepideh H. Effect of mycorrhizal fungi on some physiological characteristics of salt stressed *Ocimum basilicum L*. Iraninan Journal of Plant Physiology 2011;1 (4):215–22.
- Evelin H, Kapoor R, Giri B. Arbuscular mycorrhizal fungi in alleviation of salt stress: a review. Annals of Botany 2009;104:1263–80.
- 135. Augé RM, Toler HD, Saxton AM. Arbuscular mycorrhizal symbiosis alters stomatal conductance of host plants more under drought than under amply watered conditions: a meta-analysis. Mycorrhiza 2014;25(1):13–24.
- Kapoor R, Sharma D, Bhatnagar AK. Arbuscular mycorrhizae in micropropagation systems and their potential applications. Scientia Horticulturae 2008;116:227–39.
- 137. Kaya C, Ashraf M, Sonmez O, Aydemir S, Tuna AL, Cullu MA. The influence of arbuscular mycorrhizal colonisation on key growth parameters and fruit yield of pepper plants grown at high salinity. Scientia Horticulturae 2009;121:1–6.
- Augé RM, Schekel KA, Wample RL. Leaf water and carbohydrate status of VA mycorrhizal rose exposed to drought stress. Plant and Soil 1987;99:291–302.
- Jeffries P, Barea JM. Arbuscular mycorrhiza: a key component of sustainable plant–soil ecosystems. In: Hock B, editor. The Mycota. Vol IX: Fungal Associations. Springer-Verlag, Berlin, Germany; 2001. p. 95–113.
- 140. Rivero RM, Mestre TC, Mittler R, Rubio F, Garcia-Sanches F, Martinez V. The combined effect of salinity and heat reveals a specific physiological, biochemical and molecular response in tomato plants. Plant, Cell and Environment 2014;37:1059–73.
- Ruiz-Lozano JM. Arbuscular mycorrhizal symbiosis and alleviation of osmotic stress. New perspectives for molecular studies. Mycorrhiza 2003;13:309–17.
- 142. Fitter A. Why plant science matters. New Phytologist 2012;193(1):1–2.
- 143. Marulanda-Aguirre A, Azcon R, Ruiz-Lozano JM, Aroca R. Differential effects of a *Bacillus megaterium* strain on Lactuca sativa plant growth depending on the origin of the arbuscular mycorrhizal fungus coinoculated: physiologic and biochemical traits. Journal of Plant Growth Regulation 2008;27:10–18.

- 144. Marulanda A, Barea JM. Stimulation of plant growth and drought tolerance by native microorganisms (AMF and bacteria) from dry environments: mechanisms related to bacterial effectiveness. Journal of Plant Growth Regulation 2009;28:115–24.
- Al-Karaki GN, McMichael B, Zak J. Field response of wheat to arbuscular mycorrhizal fungi and drought stress. Mycorrhiza 2004;14:263–9.
- 146. Zou Y, Srivastava AK, Wu Q, Huang Y. Glomalin-related soil protein and water relations in mycorrhizal citrus (*Citrus tangerina*) during soil water deficit. Archives of Agronomy and Soil Science 2014;60(8):1103–14.
- 147. Allen MF. Mycorrhizal fungi: highways for water and nutrients in arid soils. Vadose Zone Journal 2007;6:291–7.
- 148. Aroca R, Porcel R, Ruiz-Lozano JM. How does arbuscular mycorrhizal symbiosis regulate root hydraulic properties and plasma membrane aquaporins in *Phaseolus vulgaris* under drought, cold or salinity stresses? New Phytologist 2007;173:808–16.
- Selvaraj T, Chelleppan P. Arbuscular mycorrhizae: a diverse personality. Central European Journal of Agriculture 2006;7:349–58.
- 150. Aranda E, Scervino JM, Godoy P, Reina R, Ocampoa JA, Wittich RM, *et al.* Role of arbuscular mycorrhizal fungus *Rhizophagus custos* in the dissipation of PAHs under root-organ culture conditions. Environmental Pollution 2013;181:182–9.
- 151. Khalil SE, Hussein MM, Khalil AM. Interaction effects of different soil moisture levels, arbuscular mycorrhizal fungi and three phosphate levels on: II-mineral ions, protein and amino acids contents of Garden Cress (*Lepidium sativum L.*) plant. International Journal of Advanced Research 2014;2(12):263–78.
- Gutjahr C, Paszkowski U. Multiple control levels of root system remodelling in arbuscular mycorrhizal symbiosis. Frontiers in Plant Science 2013;4(204):1–8.
- Xu H, Cooke JEK, Zwiazek JJ. Phylogenetic analysis of fungal aquaporins provides insight into their possible role in water transport of mycorrhizal associations. Botany 2013;91 (8):495–504.
- 154. Khalvati MA, Hu Y, Mozafar A, Schmidhalter U. Quantification of water uptake by arbuscular mycorrhizal hyphae and its significance for leaf growth, water relations, and gas exchange of barley subjected to drought stress. Plant Biology 2005;7:706–12.
- 155. Porcel R, Aroca R, Azcón R, Ruiz-Lozano JM. PIP aquaporin gene expression in arbuscular mycorrhizal Glycine max and Lactuca sativa plants in relation to drought stress tolerance. Plant Molecular Biology 2006;60:389–404.
- 156. Aroca R, Vernieri P, Ruiz-Lozano JM. Mycorrhizal and non-mycorrhizal Lactuca sativa plants exhibit contrasting responses to exogenous ABA during drought stress and recovery. Journal of Experimental Botany 2008b;59:2029–41.
- 157. Lee SH, Calvo-Polanco M, Chung GC, Zwiazek JJ. Cell water flow properties in root cortex of ectomycorrhizal (Pinus banksiana) seedlings. Plant, Cell and Environment 2010;33:769–80.
- Mack RN, Simberloff D, Lonsdale WM, Evans H, Clout M, Bazzaz FA. Biotic invasions: causes, epidemiology, global consequences, and control. Ecological Applications 2010;10:689–710.

- 159. Vaast P, Caswell-Chen EP, Zasoski RJ. Influences of a root-lesion nematode, Pratylenchus coffeae, and two arbuscular mycorrhizal fungi, Acaulospora mnellea and Glomus clarum, on coffee (*Coffea arabica L.*). Biology and Fertility of Soils 1998;26:130–5.
- Larsen J, Yohalem D. Interactions between mycorrhiza and powdery mildew of cucumber. Mycological Progress 2004;3(2):123–8.
- Raaijmakers JM, Paulitz TC, Steinberg C, Alabouvette C, Moenne-Loccoz Y. The rhizosphere: a playground and battlefield for soilborne pathogens and beneficial microorganisms. Plant and Soil 2009;321:341–61.
- 162. Pozo MJ, Slezack-Deschaumes S, Dumas-Gaudot E, Gianinazzi S, Azcón-Aguilar C. Plant defense responses induced by arbuscular mycorrhizal fungi. In: Gianinazzi S, Schuepp H, Barea JM, editors. Mycorrhizal Technology in Agriculture. Birkhäuser Verlag, Basel, Switzerland; 2002. p. 103–12.
- 163. Abdel-Fattah GM, El-Habbad SA, Hafez EE, Rashadd YM. Induction of defense responses in common bean plants by arbuscular mycorrhizal fungi. Microbiological Research 2011;166:268–81.
- Norman JR, Hooker JE. Sporulation of *Phytophthora fragariae* shows greater stimulation by exudates of non-mycorrhizal than by mycorrhizal strawberry roots. Mycological Research 2000;104:1069–73.
- 165. Liu J, Maldonado-Mendoza I, Lopez-Meyer M, Cheung F, Town CD, Harrison MJ. Arbuscular mycorrhizal symbiosis is accompanied by local and systemic alterations in gene expression and an increase in disease resistance in the shoots. Plant Journal 2007;50:529–44.
- 166. Badri DV, Vivanco JM. Regulation and function of root exudates. Plant, Cell and Environment 2009;32:666–81.
- 167. Slezack S, Dumas-Gaudot E, Paynot M, Gianinazzi S. Is a fully established arbuscular mycorrhizal symbiosis required for bioprotection of Pisum sativum roots against *Aphanomyces euteiches*? Molecular Plant-Microbe Interactions 2000;13:238–41.
- Wehner J, Autunes PM, Powell JR, Mazukatow J, Rillig MC. Plant pathogen protection by arbuscular mycorrhizas: a role for fungal diversity? Pedobiologia 2010;53(3):197–201.
- 169. Van der Heijden MGA, Streitwolf-Engel R, Riedl R, Siegrist S, Neudecker A, Ineichen K, *et al.* The mycorrhizal contribution to plant productivity, plant nutrition and soil structure in experimental grassland. New Phytologist 2006;172:739–52.
- Bardgett RD. The Biology of Soil. A Community and Ecosystem Approach. Oxford University Press, Oxford, UK; 2005.
- Pineda A, Soler R, Pozo MJ, Rasmann S, Turlings TCJ. Above-belowground interactions involving plants, microbes and insects. Frontiers in Plant Science 2015;6(318):1–3.
- 172. Van Der Putten WH, Vet LEM, Harvey JA, Wäckers FL. Linking above- and below-ground multitrophic interactions of plants, herbivores, pathogens, and their antagonists. Trends in Ecology and Evolution 2001;16:547–54.
- 173. Bennett AE, Alers-Garcia J, Bever JD. Three-way interactions among mutualistic mycorrhizal fungi, plants, and plant enemies: hypotheses and synthesis. American Naturalist 2006;167(2):141–52.

- 174. Biere A, Bennett AE. Three-way interactions between plants, microbes and insects. Functional Ecology 2013;27:567–73.
- 175. Gilbert L, Johnson D. Plant-mediated 'apparent effects' between mycorrhiza and insect herbivores. Current Opinion in Plant Biology 2015;26:100–5.
- 176. Tao L, Ahmad A, Roode JC, Hunter MD. Arbuscular mycorrhizal fungi affect plant tolerance and chemical defences to herbivory through different mechanisms. Journal of Ecology 2016;104:561–71.
- 177. Pozo MJ, Verhage A, García-Andrade J, García JM, Azcón-Aguilar C. Priming plant defences against pathogens by arbuscular mycorrhizal fungi. In: Azcón-Aguilar C, Barea JM, Gianinazzi S, Gianinazzi-Pearson V, editors. Mycorrhizas: Functional Processes and Ecological Impact. Springer, Heidelberg, Germany; 2009. p. 123–36.
- 178. Campos-Soriano L, García-Martínez J, Segundo BS. The arbuscular mycorrhizal symbiosis promotes the systemic induction of regulatory defense-related genes in rice leaves and confers resistance to pathogen infection. Molecular Plant Pathology 2012;13(6):579–92.
- Barto EK, Rillig MC. Does herbivory really suppress mycorrhiza? A meta-analysis. Journal of Ecology 2010;98(4):745–53.
- Koricheva J, Gange AC, Jones T. Effects of mycorrhizal fungi on insect herbivores: a meta-analysis. Ecology 2009;90:2088–97.
- Yang H, Dai Y, Wang X, Zhang Q, Zhu L, Bian X. Meta-analysis of interactions between Arbuscular Mycorrhizal fungi and biotic stressors of plants. Scientific World Journal 2014;746506. p. 1–7.
- 182. Gianinazzi S, Gianinazzi-Pearson V. Mycorrhizae: a plant's health insurance. Chimica Oggi 1988;10:56–68.
- Mukerji KG, Ciancio A. Mycorrhizae in the integrated pest and disease management. In: Ciancio A, Mukerji KG, editors. General concepts in integrated pest and disease management. Springer, Dordrecht, Netherlands; 2007. p. 245–66.
- Cruz AF, de Oliveira Soarez WD, Blum LEB. Impact of the arbuscular mycorrhizal fungi and bacteria on biocontrol of White Root Rot in fruit seedlings. Journal of Plant Physiology and Pathology 2014;2(1):1–5.
- 185. Mosquera-Espinosa AT, Bayman P, Prado GA, Gómez-Carabalí A, Otero JT. The double life of Ceratobasidium: orchid mycorrhizal fungi and their potential for biocontrol of Rhizoctonia solani sheath blight of rice. Mycologia 2013;105(1):141–15.
- 186. Vos CM, Tesfahun AN, Panis B, De Waele D, Elsen A. Arbuscular mycorrhizal fungi induce systemic resistance in tomato against the sedentary nematode Meloidogyne incognita and the migratory nematode *Pratylenchus penetrans*. Applied Soil Ecology 2012a;61:1–6.
- 187. Pineda A, Dicke M, Pieterse CMJ, Pozo MJ. Beneficial microbes in a changing environment: are they always helping plants to deal with insects? Functional Ecology 2013;27:574–86.
- Dong LQ, Zhang KQ. Microbial control of plant-parasitic nematodes: a five-party interaction. Plant and Soil 2006;288:31–45.

- 189. Babikova Z, Johnson D, Bruce T, Pickett J, Gilbert L. Underground allies: how and why do mycelial networks help plants defend themselves? Bioessays 2013;36:21–6.
- 190. Kabata-Pendias A. Trace Elements in Soil and Plants. CRC Press, Boca Raton, FL, USA; 2011.
- 191. Cabral L, Sousa Soares CR, Giachini AJ, Siqueira JO. Arbuscular mycorrhizal fungi in phytoremediation of contaminated areas by trace elements: mechanisms and major benefits of their applications. World Journal of Microbiology and Biotechnology 2015;31(11):1655–64.
- 192. Prasad A, Kumar S, Khaliq A, Pandey A. Heavy metals and arbuscular mycorrhizal (AM) fungi can alter the yield and chemical composition of volatile oil of sweet basil (*Ocimum basilicum* L.). Biology and Fertility of Soils 2011;47(8):853–61.
- Lin YF, Aarts MGM. The molecular mechanism of zinc and cadmium stress response in plants. Cellular and Molecular Life Sciences 2012;69(19):3187–206.
- 194. Khan A, Khan S, Khan MA, Qamar Z, Waqas M. The uptake and bioaccumulation of heavy metals by food plants, their effects on plants nutrients, and associated health risk: a review. Environmental Science and Pollution Research 2015;22(18):13772–99.
- 195. Saba H, Jyoti P, Neha S. Mycorrhizae and Phytochelators as remedy in heavy metal contaminated land remediation. International Research Journal of Environment Sciences 2013;2(1):74–8.
- 196. Cheng S, Grosse W, Karrenbrock F, Thoennessen M. Efficiency of constructed wetlands in decontamination of water polluted by heavy metals. Ecological Engineering 2002;18(3):317–25.
- 197. Lasat HA. Phytoextraction of toxic metals: a review of biological mechanisms. Journal of Environmental Quality 2002;31(1):109–20.
- 198. Schneider J, Stürmer SL, Guimarães Guilhermea LR, de Souza Moreira FM, Fonsêca de Sousa Soares CR. Arbuscular mycorrhizal fungi in arsenic-contaminated areas in Brazil. Journal of Hazardous Materials 2013;262:1105–15.
- Daghino S, Martino E, Perotto S. Model systems to unravel the molecular mechanisms of heavy metal tolerance in the ericoid mycorrhizal symbiosis. Mycorrhiza 2015;26(4):1–12.
- 200. Hildebrandt U, Regvar M, Bothe H. Arbuscular mycorrhiza and heavy metal tolerance. Phytochemistry 2007;68:139–46.
- 201. Audet P, Charest C. Effects of AM colonization on 'wild tobacco' plants grown in zinc-contaminated soil. Mycorrhiza 2006;16:277–83.
- 202. Vodnik, D, Grčman H, Maček I, van Elteren JT, Kovačevič M. The contribution of glomalin-related soil protein to Pb and Zn sequestration in polluted soil. Science of the Total Environment 2008;392(1):130–6.
- 203. González-Chávez MC, Carrillo-González R, Wright SF, Nichols KA. The role of glomalin, a protein produced by mycorrhizal fungi, in sequestering potentially toxic elements. Environmental Pollution 2004b;130:317–23.
- 204. Garg N, Chandel S. Role of arbuscular mycorrhizal (AM) fungi on growth, cadmium uptake, osmolyte, and phytochelatin synthesis in Cajanus cajan (L.) Millsp. under NaCl and Cd stresses. Journal of Plant Growth Regulation 2012;31:292–308.
- 205. Jentschke G, Goldbold DL. Metal toxicity and ectomycorrhizas. Physiologia Plantarum 2000;109:107–16.

- 206. Göhre V, Paszkowski U. Contribution of the arbuscular mycorrhizal symbiosis to heavy metal phytoremediation. Planta 2006;223(6):1115–22.
- 207. Christie P, Li X, Chen B. Arbuscular mycorrhiza can depress translocation of zinc to shoots of host plants in soils moderately polluted with zinc. Plant and Soil 2004;261:209–17.
- Joner EJ, Briones R, Levyal C. Metal-binding capacity of arbuscular mycorrhizal mycelium. Plant and Soil 2000;226:227–34.
- 209. Andrade SAL, Silveira APD, Mazzafera P. Arbuscular mycorrhiza alters metal uptake and the physiological response of *Coffea arabica* seedlings to increasing Zn and Cu concentrations in soil. Science of the Total Environment 2010;408(22):5381–91.
- 210. Giasson P, Jaouich A, Gagné S, Moutoglis P. Arbuscular mycorrhizal fungal involvement in zinc and cadmium speciation change and phytoaccumulation. Remediation 2005;15:75–81.
- 211. Ban Y, Xu Z, Zhang H, Chen H, Tang M. Soil chemistry properties, translocation of heavy metals, and mycorrhizal fungi associated with six plant species growing on lead-zinc mine-tailings. Annals of Microbiology 2015;65:503–15.
- 212. Wang F, Lin X, Yin R. Heavy metal uptake by arbuscular mycorrhizas of Elsholtzia splendens and the potential for phytoremediation of contaminated soil. Plant and Soil 2005;269:225–32.
- Soares CRFS, Siqueira JO. Mycorrhiza and phosphate protection of tropical grass species against heavy metal toxicity in multi-contamined soil. Biology and Fertility of Soils 2008;44:833–41.
- Whitfield L, Richards AJ, Rimmer DL. Relation-ships between soil heavy metal concentration and mycorrhizal colonization in *Thymus polytrichus* in northern England. Mycorrhiza 2004;14:55–62.
- 215. Adewole MB, Awotoye OO, Ohiembor MO, Salami AO. Influence of mycorrhizal fungi on phytoremediation potential and yield of sunflower in Cd and Pb polluted soils. Journal of Agricultural Sciences 2010;55(1):17–28.
- Zarei M, Hempel S, Wubet T, Schäfer T, Savaghebi G, Jouzani GS, *et al.* Molecular diversity of arbuscular mycorrhizal fungi in relation to soil chemical properties and heavy metal contamination. Environmental Pollution 2010;158(8):2757–65.
- Zarei M, Saleh-Rastin N, Jouzani GS, Savaghebi G, Buscot F. Arbuscular mycorrhizal abundance in contaminated soils around a zinc and lead deposit. European Journal of Soil Biology 2008b;44:381–91.
- Pawlowska TE, Charvat I. Heavy-metal stress and developmental patterns of arbuscular mycorrhizal fungi. Applied and Environmental Microbiology 2004;70:6643–9.
- Karimi A, Khodaverdiloo H, Sepehri M, Sadaghiani MR. Arbuscular mycorrhizal fungi and heavy metal contaminated soils. African Journal of Microbiology Research 2011;5(13):1571–6.
- 220. Gaur A, Adholeya A. Prospects of arbuscular mycorrhizal fungi in phytoremediation of heavy metal contaminated soils. Current Science 2004;86:528–34.
- 221. Khade SW, Adholeyavan A. Arbuscular mycorrhizal association in plants growing on metal-contaminated and

noncontaminated soils. Water, Air and Soil Pollution 2009;202:45–56.

- 222. Nasim G, Bajwa R. Arbuscular mycorrhizal symbiosis of wheat and exposure to air pollutants in urban Pakistan. Canadian Journal of Pure and Applied Sciences 2008;2(1):155–74.
- 223. Olsson O, Olsson PA, Hammer EC. Phosphorus and carbon availability regulate structural composition and complexity of AM fungal mycelium. Mycorrhiza 2014;24:443–51.
- 224. Thornley JHM, Parsons AJ. Allocation of new growth between shoot, root and mycorrhiza in relation to carbon, nitrogen and phosphate supply: teleonomy with maximum growth rate. Journal of Theoretical Biology 2014;342:1–14.
- 225. Hodge A. The plastic plant: root responses to heterogeneous supplies of nutrients. New Phytologist 2004;162:9–24.
- 226. Andersen CP. Source-sink balance and carbon allocation below ground in plants exposed to ozone. New Phytologist 2003;157:213–28.
- 227. Veresoglou SD, Menexes G, Rillig MC. Do arbuscular mycorrhizal fungi affect the allometric partition of host plant biomass to shoots and roots? A meta-analysis of studies from 1990 to 2010. Mycorrhiza 2012;22:227–35.
- 228. Koide RT. Functional complementarity in the arbuscular mycorrhizal symbiosis. New Phytologist 2000;147(2):233–5.
- Koide RT. Mycorrhizal symbiosis and plant reproduction. In: Koltai H, Kapulnik Y, editors. Arbuscular Mycorrhizas: Physiology and Function. 2nd ed. Springer, Dordrecht, Netherlands; 2010. p. 297–320.
- Baum C, El-Tohamy W, Grunda N. Increasing the productivity and product quality of vegetable crops using arbuscular mycorrhizal fungi: a review. Scientia Horticulturae 2015;187:131–41.
- 231. FAO. The State of Food Insecurity in the World in 2010: Addressing Food Insecurity in Protracted Crises. FAO, Rome, Italy; 2010b.
- 232. Sanchez PA. Ecology soil fertility and hunger in Africa. Science 2002;295:2019–20.
- 233. FAO. The State of Food Insecurity in the World UN Food and Agriculture Organization. FAO, Rome, Italy; 2008.
- Altieri MA. The ecological role of biodiversity in agroecosystems. Agriculture, Ecosystems and Environment 1999;74:19–21.
- 235. Hart MM, Trevors JT. Microbe management: application of mycorrhizal fungi in sustainable agriculture. Frontiers in Ecology and the Environment 2005;3:533–9.
- 236. Gianinazzi S, Gollotte A, Binet M-N, van Tuinen D, Redecker D, Wipf D. Agroecology: the key role of arbuscular mycorrhizas in ecosystem services. Mycorrhiza 2010;20(8):519–30.
- 237. Oehl F, Sieverding E, Ineichen K, Ris EA, Boller T, Wiemken A. Community structure of arbuscular mycorrhizal fungi at different soil depths in extensively and intensively managed agroecosystems. New Phytologist 2005;165:273–83.
- 238. Mozafar A, Anken T, Ruh R, Frossard E. Tillage intensity, mycorrhizal and nonmycorrhizal fungi, and nutrient concentrations in maize, wheat, and canola. Agronomy Journal 2000;92:1117–24.
- 239. Karasawa T, Takebe M. Temporal or spatial arrangements of cover crops to promote arbuscular mycorrhizal colonization

and P uptake of upland crops grown after nonmycorrhizal crops. Plant and Soil 2011;353:355–66.

- 240. Schnoor TK, Lekberg Y, Rosendahl S, Olsson PA. Mechanical soil disturbance as a determinant of arbuscular mycorrhizal fungal communities in semi-natural grassland. Mycorrhiza 2011;21:211–20.
- Hijri I, Sykorova Z, Oehl F, Ineichen K, Mäder P, Wiemken A, et al. Communities of arbuscular mycorrhizal fungi in arable soils are not necessarily low in diversity. Molecular Ecology 2006;15:2277–89.
- 242. Déry P, Anderson B. Peak phosphorus. Energy Bulletin [serial online] 2007. August 13 2007. Available from: URL: http://www. resilience.org/stories/2007-08-13/peak-phosphorus.
- 243. Lal R. Restoring soil quality to mitigate soil degradation. Sustainability 2015;7:5875–95.
- Godfray HC, Garnett T. Food security and sustainable intensification. Philosophical Transactions of the Royal Society B 2014;369(1639):20120273.
- 245. Lal R. Soil degradation as a reason for inadequate human nutrition. Food Security 2009;1:45–57.
- Bethlenfalvay GJ, Linderman RG (editors). Mycorrhizae in Sustainable Agriculture. American Society of Agronomy, Madison, WI, USA; 1992. Special publication no 54.
- De Schutter O, Vanloqueren G. The new green revolution: how twenty-first-century science can feed the world. Solutions 2011;2(4):33–44.
- 248. Garnett T, Appleby MC, Balmford A, Bateman IJ, Benton TG, Bloomer P, *et al.* Sustainable intensification in agriculture: premises and policies. Science 2013;341:33–4.
- Montanarella L, Vargas R. Global governance of soil resources as a necessary condition for sustainable development. Current Opinion in Environmental Sustainability 2012;4(5):559–64.
- 250. Fleskens L, Stringer LC. Land management and policy responses to mitigate desertification and Land degradation. Land Degradation and Development 2014;25(1):1–4.
- 251. Whipps JM. Prospects and limitations for mycorrhizas in biocontrol of root pathogens. Can Journal of Botany 2004;82:1198–227.
- 252. Leifheit E. Soil sustainability and arbuscular mycorrhizal fungi [PhD thesis]. Freie UniversitätBerlin, Berlin; 2014.
- 253. Bardgett RD, Bowman WD, Kaufmann R, Schmidt SK. A temporal approach to linking aboveground and belowground ecology. Trends in Ecology and Evolution 2005;20:634–41.
- 254. Six J, Bossuyt H, Degryze S, Denef K. A history of research on the link between (micro)aggregates, soil biota, and soil organic matter dynamics. Soil Tillage Research 2004;79:7–31.
- 255. Poorter H, Buehler J, van Dusschoten D, Climent J, Postma JA. Pot size matters: a meta-analysis of the effects of rooting volume on plant growth. Functional Plant Biology 2012;39:839–50.
- 256. Piotrowski JS, Denich T, Klironomos JN, Graham JM, Rillig MC. The effects of arbuscular mycorrhizas on soil aggregation depend on the interaction between plant and fungal species. New Phytol 2004;164:365–73.
- 257. Sanchez PA, Ahamed S, Carré F, Hartemink AE, Hempel J, Huising J, *et al.* Digital soil map of the world. Science 2009;325:680–1.

- 258. Hendrey GR. Global greenhouse studies: need for a new approach to ecosystem manipulation. Critical Reviews in Plant Science 1992;11:61–74.
- 259. Cameron DD. A role for differential host resistance to the hemiparasitic angiosperm Rhinanthus minor L. in determining the structure of host plant communities? [PhD thesis]. University of Aberdeen, Aberdeen, UK; 2004.
- Gehring CA, Whitham TG. Mycorrhizae-herbivore interactions: population and community consequences. In: Van Der Heijden MGA, Sanders IR, editors. Mycorrhizal Ecology. Springer, Berlin, Germany; 2002. p. 295–320.
- 261. Gehring CA, Bennett AE. Mycorrhizal fungal-plant-insect interactions: the importance of a community approach. Environmental Entomology 2009;38:93–102.

- Kardol P, De Deyn GB, Laliberté E, Mariotte P, Hawkes CV. Biotic plant–soil feedbacks across temporal scales. Journal of Ecology 2013;101(2):309–15.
- 263. Vanderlinden K, Vereecken H, Hardelauf H, Pachepsky Y. Temporal stability of soil water contents: a review of data and analyses. Vadose Zone Journal 2012;11(4):19.
- 264. Wardlaw LA. The control of carbon partitioning in plants. New Phytologist 1990;116:341–81.
- 265. Staddon PL, Heinemeyer A, Fitter AH. Mycorrhizas and global environmental change: research at different scales. Plant and Soil 2002;244:253–61.
- 266. Koide RT. Context-Dependent Interaction Hierarchies and the Organization of Ectomycorrhizal Fungal Communities. In: Hock B, editor. The Mycota, Volume IX: Fungal Associations. Springer-Verlag, Berlin; 2012. p. 181–95.